

An Overview of the Use of Fatty Acids in Fish Farming Research during the Last Decade, with Particular Emphasis on Fish Quality

NAFSIKA KARAKATSOULI

Department of Applied Hydrobiology, Faculty of Animal Science and Aquaculture, Agricultural University of Athens, 118 55 Athens, Greece

Abstract

The present literature overview was carried out in order to assess the use of fatty acids (FA) analysis in fish farming research during the last decade. As aquaculture products are mainly intended for human consumption, the overview is focused on fish fillet, carcass or whole body FA, as a major aspect of fish nutritional value. This survey used 394 papers and demonstrated that a) 71% of papers report FA as relative percentage of total FA (% totFA), b) 23% of papers report actual FA amounts, c) in 63% of papers not enough information is provided in "Materials and Methods" section to understand exactly how FA analysis was performed, and d) based on FA percentages, results are not always the same as when based on FA amounts, and thus divergent conclusions about treatment effects could be drawn. These findings indicate that certain issues need to be addressed regarding adequate description of analytical methods followed for FA analysis, presentation and interpretation of FA results and may help to highlight critical points that should be considered from the researchers' and scientific journals' sides in order to foster more informative research publications.

As fish consumption was linked with reduced incidence of cardiovascular disease in the Greenland Eskimos (Dyerberg et al. 1975; Givens and Gibbs 2008), increasing evidence helped to further associate long-chain omega-3 (n-3) polyunsaturated fatty acids (LC n-3 PUFA) with several human health benefits (Ruxton and Derbyshire 2009). The major dietary source of the most beneficial LC n-3 PUFA (eicosapentaenoic acid EPA, 20:5n-3; docosapentaenoic acid DPA, 22:5n-3; docosahexaenoic acid DHA, 22:6n-3) is seafood. This fact explains the increased research interest in aquaculture products, farmed fish in particular. Research is mainly focused on understanding fish fatty acids (FA) biochemical and metabolic pathways, as well as on finding ways to improve LC n-3 PUFA content of the product that reaches the consumer.

It is common knowledge that nutritional value of a certain food is deduced by its nutrient content (i.e., nutrient amount). In the case of fish, it is the fish flesh high amounts of LC n-3 PUFA (among other nutrients) that render fish a food of high nutritional value. It

is also common knowledge that fish farming industry produces fish for human consumption and is responsible for producing a high-quality product. It is reasonable to assume that when data about farmed fish flesh FA absolute amount is needed, this data would be found in fish farming (aquaculture) related journals. However, a simple search immediately shows that the relevant information is rather hard to find, despite the great number of research papers that report FA analysis. Instead, much more related information can be found in papers of "Food Science" journals, even if these studies focus on food quality aspects after the product is harvested/slaughtered.

Motivated by the belief that it is the aquaculture sector's obligation and responsibility to produce high-quality fish, through appropriate production practices (e.g., fish nutrition, rearing conditions, protection of fish health and welfare etc.), and in order to assess the current situation, a literature overview was carried out using research works published during the last decade in fish farming related journals. The present study reports the main findings and

TABLE 1. *Relative frequency (%; n = 394) of papers included in the overview, classified by scientific journal and publication year (January 2000–August 2010).*

Journal name (alphabetical order, n = 21)	Publication year		
Acta Ichthyologica et Piscatoria	0.51	2000	4.82
Aquacultural Engineering	0.51	2001	5.33
Aquaculture International	4.57	2002	4.57
Aquaculture Nutrition	15.23	2003	9.64
Aquaculture Research	13.20	2004	7.11
Aquaculture	37.56	2005	7.87
Aquatic Living Resources	0.51	2006	7.61
Canadian Journal of Fisheries and Aquatic Sciences	0.25	2007	12.94
Fish Physiology and Biochemistry	5.08	2008	14.97
Fish and Shellfish Immunology	0.25	2009	13.96
Fisheries Science	6.09	2010 (August)	11.17
ICES Journal of Marine Science	0.51		
Iranian Journal of Fisheries Sciences	0.25		
Journal of Applied Ichthyology	3.81		
Journal of Fish Biology	1.52		
Journal of the World Aquaculture Society	4.57		
North American Journal of Aquaculture	2.79		
North American Journal of Fisheries Management	0.51		
Reviews in Fish Biology and Fisheries	0.51		
The Israeli Journal of Aquaculture (Bamidgeh)	1.27		
Turkish Journal of Fisheries and Aquatic Sciences	0.51		

conclusions of this survey giving general statistics on the use of FA analysis in fish farming research emphasizing on fish quality. It also points out the most important issues related to description of analytical methods, FA presentation and results interpretation. The aim of this work is to highlight certain points that should be considered from the researchers and scientific journals sides in order to foster more informative research publications.

Materials and Methods

Selection of Studies – Inclusion Criteria

In order to have an unbiased criterion for journal selection, scientific journals included in this overview come from the subject category “Fisheries” of the 2009 Thomson Reuters (ISI) Journal Citation Reports Ranking (47 scientific journals in total). This subject category was chosen due to a) the lack of an “Aquaculture” subject category, b) the fact that it contains aquaculture-related scientific journals, and c) the next more relevant subject category was “Marine and Freshwater Biology” which included only 16 out of the 47 journals of

the selected category. A thorough search was performed in all 47 journals. Selected papers had to fulfill the following criteria: a) to be research papers published during the last decade (2000–August 2010), b) to include FA analysis in fish fillet (muscle), whole body or carcass, c) to involve species of interest for aquaculture (either already widely farmed or presenting strong potential for farming), and d) to be published in English (only 2 out of 47 journals were excluded). Three hundred ninety four (394) papers were found to meet these criteria (Appendix), published in 21 Journals (Table 1). Full texts of all 394 papers were obtained and assessed.

Information Recorded – Classifications – Definitions

For each paper several parameters were recorded and used for further analysis. The chosen parameters aimed to obtain a) general information of selected papers fulfilling the above-mentioned criteria (e.g., time-course of research, species used, treatment etc.) and b) specific information as to how and why FA analysis is performed in relation to the way

FA results are presented (e.g., description of methods used, objective, presentation).

Information recorded was defined and classified as follows:

Publication year: It refers to the year of publication and not of acceptance. Also, for the year 2010 all online-first related papers searched till the 31st August 2010 were included.

Country of origin where research was carried out: Almost all selected papers had more than one author. To have an indication of where the main part of research related to FA was carried out, the country of origin of the corresponding author, also assumed to be the researcher in charge of the presented work, was recorded.

Species used: The scientific name of species used was recorded. It was also taken into account that some papers studied more than one species.

Fish life stage – body weight: It refers to life stage – body weight at sampling time of tissues for FA analysis.

Experimental treatment: According to experimental treatment, selected papers were divided into four groups: papers a) studying fish nutrition, b) studying the effects of several rearing conditions, c) aiming to compare wild versus farmed fish, and d) papers classified as “others” which included all other treatments encountered of lower frequencies.

Tissue and lipid class analyzed for FA: Apart from classifying papers according to whether fillet (muscle), whole body or carcass was used for FA analysis (inclusion criteria b), the use of total lipid or lipid classes (e.g., triacylglycerides, phospholipids etc.) was also recorded.

Objective of FA analysis: Selected papers were divided into three major groups according to what the authors stated to be their purpose for FA analysis, or else if this was not clearly declared, according to how the authors used FA results for drawing their conclusions. These include papers a) using FA from a strictly physiological point of view (hereafter named “Physiology”), b) reporting FA as a parameter of fish quality (hereafter named “Quality”), and

c) using FA to conclude on both physiological aspects and quality (hereafter named “Both”).

Presentation of FA results: Three major groups were formed. The first group (hereafter named “Percentage”) includes papers that report FA as percentage (area % or weight, wt %) of totFA (i.e., relative proportion of each FA among totFA) and do not report totFA content of tissue analyzed. The second group (hereafter named “Amount”) includes papers that report actual FA content or give enough information to calculate FA amounts. Units encountered were a) weight of FA per weight of tissue analyzed (wt/wt) on a wet or dry basis, b) weight of FA per weight of lipid analyzed (wt/wt) and lipid content of tissue is also reported, c) weight of FA per larva, in some of the papers studying this life stage, and even d) wt % of totFA, but in this case totFA content of tissue is reported. The third group (hereafter named “Amount and Percentage”) includes papers that report both ways of expression.

Description of methods used for FA analysis: The main criterion for this parameter was whether the authors report enough information to understand all analytical steps involved in FA analysis (i.e., lipid extraction, FA methylation and FA chromatographic analysis: peak separation, identification, and quantification).

Evaluation of Information Recorded

For each of the above mentioned parameters, frequency tables were constructed. In addition, in order to obtain a deeper insight of recorded information, cross tabulations among recorded parameters were also constructed. In all tables relative frequency (as percentage of total number of papers) is reported. In the text, frequency (absolute number of papers) is also sometimes reported. Classes with lower frequencies than those presented are pooled and reported under the name “Others.”

Results

General Information for Fish FA Research

During the last decade there is a gradual and steady increase of research papers concerning farmed fish FA (Table 1), papers published

TABLE 2. Relative frequency (%; n = 394) of selected papers classified by geographical origin (continent) and major countries (top 10 out of 35).

Continent	Country		
Africa	1.02	USA	11.42
Asia	23.10	Spain	10.66
Australia	5.08	Norway	10.15
Europe	50.51	Japan	8.38
North America	18.53	Canada	6.09
South America	1.78	UK	5.58
		Australia	5.08
		Italy	4.57
		France	4.06
		China	3.55
		Others (n = 25) ^a	30.46

^a“Others” include (alphabetical order): Belgium, Brazil, Chile, Croatia, Czech Republic, Denmark, Egypt, Germany, Greece, Hungary, India, Iran, Israel, Kenya, Korea, Malaysia, Mexico, Netherlands, Philippines, Poland, Portugal, Sweden, Taiwan, Tunisia and Turkey.

over the last 4 yr, that is, 2007–2010, representing 53.04% of selected studies. In about 50% of the 394 papers assessed, research took place in Europe, followed by Asia and North America (Table 2). However, the leading country in fish FA research is USA, closely followed by Spain and Norway (Table 2). Research performed in the top 10 out of 35 countries (274 studies out of 394) constituted 69.54% of total selected studies (Table 2). Despite the large number of species involved (110 in total), papers investigating FA in only 12 of them make up 53.91% (Table 3). The most studied species are Atlantic salmon and rainbow trout (Salmonidae), Atlantic cod, the two widely reared Mediterranean species gilthead sea bream and European sea bass, and tilapias (Table 3). The majority of selected papers use juvenile fish, followed by larvae and fish of commercial size (Table 4). Nutritional treatments exceed by far all other experimental factors investigated accounting for 78.14% of the studies (Table 5). Next in rank come the investigation of rearing conditions (e.g., density, temperature, stress etc.) and comparisons between farmed species and their wild conspecifics (Table 5). More than half of the selected papers report FA in fish fillet (224 studies out of 394, 56.85%), while the rest use whole body (154

TABLE 3. Relative frequency (%; n = 408^a) of studied species (top 12 out of 110) in selected papers.

Species	
Atlantic salmon, <i>Salmo salar</i>	9.80
Gilthead sea bream, <i>Sparus aurata</i>	8.33
Rainbow trout, <i>Oncorhynchus mykiss</i>	7.11
Atlantic cod, <i>Gadus morhua</i>	4.90
European sea bass, <i>Dicentrarchus labrax</i>	4.41
Tilapias, <i>Oreochromis</i> sp.	4.17
Atlantic halibut, <i>Hippoglossus hippoglossus</i>	2.94
Common carp, <i>Cyprinus carpio</i>	2.45
Channel catfish, <i>Ictalurus punctatus</i>	2.45
Murray cod, <i>Maccullochella peelii peelii</i>	2.45
Sunshine bass, <i>Morone chrysops</i> x <i>M. saxatilis</i>	2.45
Pike perch, <i>Sander lucioperca</i>	2.45
Others (n = 98)	46.09

^aOf 394 papers, 14 were included in the overview, studied more than one species.

TABLE 4. Relative frequency (%; n = 395^a) of fish life stage on sampling in selected papers.

Fish life stage	
Juvenile	45.32
Larvae	27.59
Final (commercial size)	21.52
Others (fry, parr, parr-smolt, smolt, fingerlings)	5.57

^aOf 394 papers, one was included in the overview, studied more than one life stage.

TABLE 5. Relative frequency (%; n = 430^a) of experimental treatment of selected papers.

Experimental treatment	
Nutritional	78.14
Rearing conditions	10.46
Wild versus farmed	3.95
Others ^b	7.45

^aOf 394 papers, 36 were included in the overview, investigated the effects of more than one treatment.

^b“Others” include: Ontogeny and embryonic development, processing or assessment of final product, reproduction, seasonal variations, genetics, comparison among species or tissues, fatty acid biochemistry-metabolic pathways.

out of 394, 39.09%) and only few fish carcass (16 out of 394, 4.06%).

Objective of FA Analysis

In all 394 papers, the answer to why FA analysis was performed and reported was easy

TABLE 6. *Relative frequency (%) of selected papers classified by objective, presentation and clarity of description of FA (fatty acids) analysis.*

Classification	
Objective of FA analysis	
Physiology	52.03
Quality	18.53
Both	29.44
Presentation of FA analysis	
Percentage	71.32
Amount	23.10
Both (percentage and amount)	5.08
Ratios	0.51
Clarity of description of FA analysis ^a	
Clear	36.81
“...identified...”	14.47
Detailed description of quantification method	13.71
Area	6.35
External laboratory	1.52
Direct comparison	0.76
Unclear	63.19
Not enough information	42.89
Internal standard without usage specification	20.30

^aRefer to text for details on subgroups used.

to identify and classify (Table 6). The majority of papers used FA composition as a measure for evaluating a treatment effect on several physiological issues, for example, fish health, reproduction, lipid metabolism etc. The use of FA composition to exclusively evaluate fish quality was dealt in three times less papers, while the remaining papers reported FA in order to assess both physiological status and quality.

Presentation of FA Analysis

According to the definitions presented in the previous section, in the overwhelming majority of papers FA are reported as Percentage (Table 6). In 91 out of 394 papers FA are reported as Amount and in only 20 both expressions are used. There are also two papers reporting ratios of selected FA or of sums of FA.

Description of Methods Used for FA Analysis

Generally there was adequate description of lipid extraction and methylation procedures or reference to well-documented papers or official methods (Folch et al. 1957; Christie 1990;

AOAC 2000). To complete FA analysis almost all papers report using gas chromatography (385 out of 394, 97.72%). One paper reports using HPLC (0.25%). In four papers out of 394 it was not possible to confirm the analytical method: in one of them authors state that standard methods were followed but they do not give any reference (Rosenlund et al. 2001), while in the other three the authors cite papers published in languages other than English. Also, in another four papers, the authors state that an external commercial laboratory was used for FA analysis without giving any further details. It should be noted that papers using an external laboratory are altogether six, but in two of them there is a description of the method used.

In 327 out of 394 papers (82.99%) total lipid of tissue examined was used for FA analysis. The remaining papers (67 out of 394, 17.01%) report FA of specific lipid classes (e.g., neutral and polar or triacylglycerides and phospholipids). In almost all papers (380 out of 394, 96.45%) it is clearly reported that FA were methylated and then subjected to gas chromatography. The other 14 papers (3.55%) were classified as “Unknown – Not Reported” including papers using external laboratory with no further details (4 out of 14), papers that simply do not mention the fact (5 out of 14) and papers for which the citation used was hardly accessible (5 out of 14).

Examining closely chromatographic analysis of FA as described by the authors, it should be mentioned that in almost all papers gas chromatographic conditions for FA peak separation (i.e., temperature programming and gas flow rates, column and detector used, etc.) are adequately reported either in the given paper or in the cited reference. The only exception concerns those papers that used an external laboratory with no further details (4 out of 394) or those that cite a reference in languages other than English (3 out of 394). Similarly, identification of FA is adequately described as, in almost all selected papers, authors do report that a mixture of standards or a well-characterized fish oil was used to compare peak retention times.

TABLE 7. Cross tabulation among objective, presentation, and clarity of description of FA (fatty acids) analysis.^a

Objective	Presentation	Clarity of description			Row total
		Clear	Clear-details ^b	Unclear	
Physiology (Ph)	Percentage (%)	10.91	2.79	22.59	36.29
	Amount (Am)	0.00	5.08	8.12	13.20
	% and Am	0.51	0.25	1.52	2.28
	Ratios	0.00	0.00	0.25	0.25
Quality (Q)	Percentage (%)	2.54	1.02	7.36	10.91
	Amount (Am)	0.00	1.02	4.31	5.33
	% and Am	0.00	0.51	1.52	2.03
	Ratios	0.00	0.00	0.25	0.25
Both (Ph + Q)	Percentage (%)	9.14	1.78	13.20	24.11
	Amount (Am)	0.00	1.27	3.30	4.57
	% and Am	0.00	0.00	0.76	0.76
	Ratios	0.00	0.00	0.00	0.00
Column total		23.10	13.71	63.20	100.00

^aData represent relative frequency (%; $n = 394$). For separate total data see Table 6.

^bClear-details: includes the 54 out of 394 papers (13.71%, Table 6) that describe in details FA quantification method.

As far as FA quantification is concerned the situation is more complicated. The main criterion used for the classification was whether the description provides the necessary information for a reader to understand how exactly the analysis was performed. Thus, a description could be characterized as either “clear” (145 out of 394 papers, 36.81%) or “unclear” (249 out of 394 papers, 63.19%). However, within each of these two major groups, several distinct subgroups could be formed (Table 6). With reference to the “clear” group in 25 papers, it is clearly reported that area percentages are used as they are obtained from the FA chromatogram (e.g. Nguyen et al. 2008; Sharma et al. 2010). In three papers the area of an internal standard was directly compared with those obtained from each FA (Robin and Peron 2004). In six papers the use of a commercial external laboratory is clearly declared. In 57 papers the authors reported that only identification of FA took place and, accordingly, report FA as % totFA, so one does not expect to see a quantification method (Copeman and Parrish 2002). In 54 papers the quantification method is described in details (i.e., use of internal standard and serially diluted mixtures of external standards, use of empirical or theoretical response factors for each FA, system calibration) (Bell and Dick 2004; Ludwig et al. 2008; Lund et al. 2008). Concerning the “unclear” group, in 80 papers,

the use of an internal standard is mentioned but no further explanation of this use is provided (e.g., Garduño-Lugo et al. 2007; Ramos et al. 2008; Díaz-López et al. 2009). Finally, in many of the papers (169 out of 394, 42.89%) the description of quantification is completely unclear as the only information usually provided is that FA were quantified “...by use of the xx software” or “...by integration of the peak areas” (Blanchard et al. 2008; Benedito-Palos et al. 2009; Figueiredo-Silva et al. 2010). Papers describing only FA separation and identification and reporting FA amounts are also included in this group (Jankowska et al. 2008).

Objective – Presentation – Description of Methods Combined

In order to have a deeper insight on FA research, cross tabulation among objective, presentation and clarity of description of FA analysis was formulated (Table 7). To make data presented in Table 7 more informative, the 54 papers describing quantification method in details (Table 6) were handled separately and named as “clear-details,” while all other “clear” description subgroups (91 papers, Table 6) were pooled and simply named “clear.” Several interesting comments can be highlighted from data in Table 7. First, it is further confirmed that even within each objective group presentation

TABLE 8. *Progress of selected papers, classified by objective, presentation and clarity of description of FA (fatty acids) analysis, during the last decade.^a*

Objective	Presentation ^b	Clarity of description	2000–2002	2003–2006	2007–2010
Physiology (Ph)	Percentage (%)	Clear	2.79	3.55	4.57
		Clear-details	1.02	0.51	1.27
		Unclear	3.55	6.09	12.94
	Amount (Am)	Clear	0.00	0.00	0.00
		Clear-details	1.02	1.52	2.54
		Unclear	2.28	2.54	3.30
	% and Am	Clear	0.00	0.25	0.25
		Clear-details	0.00	0.00	0.25
		Unclear	0.00	0.51	1.02
Quality (Q)	Percentage (%)	Clear	0.25	0.76	1.52
		Clear-details	0.25	0.25	0.51
		Unclear	0.76	3.05	3.55
	Amount (Am)	Clear	0.00	0.00	0.00
		Clear-details	0.00	0.25	0.76
		Unclear	0.25	1.27	2.79
	% and Am	Clear	0.00	0.00	0.00
		Clear-details	0.00	0.25	0.25
		Unclear	0.25	0.76	0.51
Both (Ph + Q)	Percentage (%)	Clear	0.76	3.81	4.57
		Clear-details	0.00	0.51	1.27
		Unclear	1.27	4.57	7.36
	Amount (Am)	Clear	0.00	0.00	0.00
		Clear-details	0.00	0.51	0.76
		Unclear	0.25	1.02	2.03
	% and Am	Clear	0.00	0.00	0.00
		Clear-details	0.00	0.00	0.00
		Unclear	0.00	0.00	0.76

^aData represent relative frequency (%; $n = 394$). Publication years were pooled, and row and column totals were omitted for simplicity.

^bPapers presenting FA analysis as ratios (2 out of 394 papers, 0.51%, Table 6) are not presented in the table but data were included in the calculation of relative frequency.

of FA as percentage of totFA is the most frequent way to report FA results (row total of Table 7). Second, papers concluding on fish quality (objective Quality and both of Table 7, 189 out of 394 studies, 47.96%) and reporting actual FA amount are 50 out of 189. Last, but not least, from those 50 papers FA quantification is described in details in just 11 (De Francesco et al. 2007; St-Hilaire et al. 2007).

Furthermore, in order to assess the progress made through the last decade, cross tabulation among all three above-mentioned parameters (i.e., objective, presentation, and clarity of description of FA analysis) and publication year was also formulated (Table 8). As publication year either handled separately or

pooled in groups of 3 and 4 yr brought about the same conclusions, to simplify data presented in Table 8 publication years were pooled (2000–2002, 3 yr, 58 out of 394 papers, 15%; 2004–2006, 4 yr, 127 out of 394 papers, 32%; 2007–2010, 4 yr, 209 out of 394 papers, 53%). From data presented in Table 8, attention can be drawn to the following points. In all Objective and Presentation groups, papers describing clearly analytical methodology used (i.e., “clear” and “clear-details” groups) have been increased from 6.09% (24 papers out of 394) over 2000–2002 to 18.52% (73 papers out of 394) over the last 4 yr. However, this threefold increase is surpassed by a fourfold increase of papers classified as “unclear” (8.63%, 34 papers

out of 394 for years 2000–2002 and 34.52%, 136 papers out of 394 for years 2007–2010). The only case that the latter fact does not hold concerns papers studying quality and reporting both FA percentage and amount, which were reduced to 0.51% over the last 4 yr from 0.76% over the years 2003–2006 (Table 8).

Discussion

The present survey aims at providing an assessment of the use of FA analysis in aquaculture-related fish studies during the last decade. Through this survey a number of issues have emerged about the justification and/or adequacy of the, up to now, use of FA. This survey emphasizes on aspects that might need improvement in order to lead to more accurate and informative scientific publications.

Presentation versus Objective of FA Analysis

The first issue concerns the way of presentation of FA results. The present survey shows clearly that presentation of FA on a percentage basis (% of totFA) is so extensively used so as to have the advantage of permitting comparisons among papers. The knowledge of the relative proportion among FA, as well as of how it may be altered because of a specific experimental treatment, is of great importance for fish homeostasis, health, and physiology. For example, papers studying fish first life stages are focused more on treatment effect on fish health and FA percentages indeed help in results evaluation (Park et al. 2006; Ludwig et al. 2008; Seychelles et al. 2011). Another example are papers focusing on dietary fish meal or fish oil replacement, mainly by plants or plant oils where FA percentages are used to evaluate farmed fish physiological status through the relative change and incorporation of FA in biological tissues (Palmegiano et al. 2006; Tidwell et al. 2007). Finally, it should not be ignored that FA percentages may involve simple calculations of chromatographic analysis (e.g., area %; Nguyen et al. 2008; Sharma et al. 2010).

However, there are also several drawbacks to the use of FA percentages. When each

FA is expressed as percentage of totFA then each FA is dependent on changes of other FA (Schwertner and Mosser 1993). In other words, when some FA are increased, one obviously expects some other FA to decrease. In cases where there is a treatment effect on totFA content, the sole use of percentages may not provide all the information that could be obtained (see below). Besides, when FA are presented as percentages without reporting actual totFA content of tissues examined, it is not possible to calculate absolute FA amounts. Lipid classes found in fish total lipids (particularly farmed fish) are triacylglycerides (and to a lesser degree diacyl- and monacyl-glycerides), phosphoglycerides, sphingolipids, cholesterol, cholesterol esters, and free fatty acids (Tocher 2003; Díaz-López et al. 2009). From these only cholesterol does not contain fatty acids. The relative contribution of these lipid classes within a specific tissue total lipid is affected by several factors such as fish species, life stage, diet, nutritional condition, physiological status etc. and thus may be subjected to differentiations due to the experimental treatment itself (Copeman and Parrish 2002; Zhu et al. 2003; Díaz-López et al. 2009). The same stands for a tissue total amount of fatty acids (Kandemir and Polat 2007). TotFA content, as percentage of total lipids, has been reported in related published papers to range from 27 to 82% (De Silva et al. 2004; De Francesco et al. 2007; Villalta et al. 2008). However, extreme values have also been reported, for example, <2% (Garduño-Lugo et al. 2007) or 100% (Kucska et al. 2006), in the latter case totFA content being calculated as if tissue total lipid all consists of FA. The use of FA as % totFA does not include possible treatment effects on FA content or on specific lipid classes (Enser et al. 1998; Hardy 2003) and thus useful information might be lost. Besides, it cannot be *per se* a measure of FA absolute amounts. Consequently, FA percentages cannot be used to conclude on fish nutritional value for human consumption. As it is very clearly stated by Hardy (2003), "...consumers do not eat percentages, they eat grams per serving." But still, 138 out of 189 of the selected papers concluding on fish

quality (Table 7), report FA solely as percentages among totFA (Ramos et al. 2008; Sharma et al. 2010). It should though be emphasized that when total lipid and FA content of the tissue analyzed is not subject to changes due to the experimental treatment, then the use of FA as % totFA is indeed illustrative of treatments effects.

On the other hand, reporting absolute FA amount has the advantage that each FA is independent of changes of other FA (Schwertner and Mosser 1993) and allows conclusions on product nutritional value (Turchini et al. 2006; De Francesco et al. 2007; Schlechtriem et al. 2007, 2009) or FA metabolic pathways (e.g., desaturation, elongation, oxidation) (Enser et al. 1998; Turchini et al. 2007). Data thus obtained can be easily used to calculate percentages if so required.

Description of Methodology for FA Analysis

Another issue that emerges from our survey is related to description of analytical methods followed for FA analysis. Although up to quantification, FA analytical procedures are adequately described (i.e., lipid extraction, FA methylation, separation, and identification), the FA quantification part seems to be the one that needs more attention, especially considering that to report FA absolute amount, FA have first to be quantified.

Principles of analytical steps that should be followed in order to quantify substances through gas chromatography have been very well reviewed and assessed (Ackman 2002; Dodds et al. 2005; Cuadros-Rodríguez et al. 2007). In brief, quantification involves peak area integration and measurement through the instrument software, system calibration, the use of internal standards and serially diluted solutions of external standards, calculation of detector response factors for each identified FA and estimation of FA recovery through the extraction and methylation procedures. In those papers, here classified as “unclear” (Table 6), the authors either state that “...the x FA was used as internal standard,” a phrase that does not imply the above-mentioned quantification

steps even if they were actually performed (e.g., Garduño-Lugo et al. 2007; Ramos et al. 2008; Díaz-López et al. 2009) or they declare that FA were quantified “...by use of the x software” or “...by integration of the peak areas” (Blanchard et al. 2008; Benedito-Palos et al. 2009; Figueiredo-Silva et al. 2010). It is, however, well known that PC software always accompanies a gas chromatogram and just the mention of a software program does not ensure the required sensitive laboratory work, which would permit the appropriate software parameters to be entered by the operator. We would like, in this point, to emphasize that an incomplete description of FA quantification does not mean that proper quantification was not performed. It just means that not enough information is provided. On the other hand, it is rather unexpected that the answer to a question like “how many papers study fish quality, report FA amount and describe in details FA quantification analysis?” is only 11 out of 189 (Table 7) (De Francesco et al. 2007; St-Hilaire et al. 2007).

It is though encouraging that during the last 4 yr there is an improvement in the number of papers that describe adequate methodology used for FA analysis and at least in one case there is a reduction of those papers here classified as describing FA analysis without analytical details (objective “Quality,” presentation “% and Am,” clarity of description “unclear,” Table 8). In contrast, it is probably disquieting that the number of papers not reporting details of methods followed increase at a greater rate.

Percentage versus Amount: Possibility of Divergent Conclusions

A final issue that reasonably follows the present survey results is whether the same conclusions can be drawn regardless of the way of FA presentation (percentage or amount). Or else, is there a possibility to obtain divergent conclusions when statistical evaluation of a treatment is based on either one of the two ways of FA presentation? The most pronounced example is the case of comparing FA of wild and farmed fish to conclude on fish

nutritional value for human consumption. It is well documented for several species that wild fish have higher EPA, DPA, DHA and total n-3 PUFA expressed as % totFA than farmed fish (Olsson et al. 2003; Blanchet et al. 2005; Jankowska et al. 2008; Sharma et al. 2010). On the basis of this fact one may draw the conclusion that wild fish are of superior nutritional value for consumers. However, as it is the absolute human requirements in n-3 PUFA that need to be covered by consuming a specific fish serving, when the lower total lipid content of wild fish is taken into account and FA are presented as amounts, then it is clear that farmed fish nutritional value is at least equal, and in most cases superior, to that of their wild conspecifics (Hardy 2003; Olsson et al. 2003; Blanchet et al. 2005). Apart from this rather evident example, there are cases where conclusions from FA percentages match those obtained from FA amounts (Bransden et al. 2005; Yildirim-Aksoy et al. 2007; Garcia et al. 2008). Nevertheless, when based on both ways of presentation valuable information might be gained that can lead to a more complete interpretation of results (e.g., see Discussion in Turchini et al. 2006; De Francesco et al. 2007; Schlechtriem et al. 2007). However, in the present survey only 20 out of the total 394 selected papers (5.08%) where FA are reported as both percentage and amount were identified.

Conclusions

The present survey demonstrated that research on farmed fish FA has been escalating during the last decade. This fact is explainable given the acknowledged importance of fish FA, especially LC n-3 PUFA, for human nutrition and health. However, despite the existing great number of papers, certain issues pertaining to analytical methodology, presentation, and discussion of FA results could be identified. Description of analytical methods followed, especially the more complicated quantification chromatographic part, should be more detailed describing thoroughly exactly how FA analysis was performed. The choice of presenting FA results as relative proportion (as % totFA),

amount or both should be carefully considered so that it justifies and better serves the aim of the given research and the initial decision to include FA as one of the analysis to be performed. Special attention is drawn to the need for reporting FA amounts in studies concluding on fish quality and nutritional value and to the fact that, in some cases, divergent conclusions could be drawn when statistics are based on percentages or on absolute FA amounts alone. Considering that fish farming industry produces fish for human consumption and that is equally concerned about fish health and growth, as well as final product quality, the use of both FA percentages and amounts when reporting FA results may be recommended. Thus, in depth information regarding FA, complete interpretation of results and sound conclusions could be obtained. Acknowledging the difficulties often encountered in lipid analysis, it is hoped that present work may initiate a fruitful discussion toward the implementation of guidelines that should limit the deficiencies outlined in this overview and improve reporting all aspects of FA analysis in scientific publications.

Acknowledgments

I am grateful to Laetitia Meyer who first introduced me in the science of chromatographic quantification techniques. Special thanks are due to my colleague K. Mountzouris for his valuable suggestions and critical reading of the manuscript. I also express my appreciation to two anonymous reviewers for their essential additions and comments.

Literature Cited

- Ackman, R. G. 2002. The gas chromatograph in practical analyses of common and uncommon fatty acids for the 21st century. *Analytica Chimica Acta* 465:175–192.
- AOAC (Association of Official Analytical Chemists). 2000. Official methods of analysis of AOAC international, 17th edition. AOAC, Gaithersburg, MD, USA.
- Bell, M. V. and J. R. Dick. 2004. Changes in capacity to synthesise 22:6n-3 during early development in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 235:393–409.
- Benedito-Palos, L., J. C. Navarro, A. Bermejo-Nogales, A. Saera-Vila, S. Kaushik, and J. Pérez-Sánchez. 2009. The time-course of fish oil wash-out follows a

- simple dilution model in gilthead sea bream (*Sparus aurata* L.) fed graded levels of vegetable oils. *Aquaculture* 288:98–105.
- Blanchard, G., J. G. Makombu, and P. Kestemont.** 2008. Influence of different dietary 18:3n-3/18:2n-6 ratio on growth performance, fatty acid composition and hepatic ultrastructure in Eurasian perch, *Perca fluviatilis*. *Aquaculture* 284:144–150.
- Blanchet, C., M. Lucas, P. Julien, R. Morin, S. Gingras, and É. Dewailly.** 2005. Fatty acid composition of wild and farmed Atlantic salmon (*Salmo salar*) and rainbow trout (*Oncorhynchus mykiss*). *Lipids* 40:529–531.
- Brandsen, M. P., S. C. Battaglene, D. T. Morehead, G. A. Dunstan, and P. D. Nichols.** 2005. Effect of dietary 22:6n-3 on growth, survival and tissue fatty acid profile of striped trumpeter (*Latris lineata*) larvae fed enriched *Artemia*. *Aquaculture* 243:331–344.
- Christie, W. W.** 1990. Preparation of methyl esters – part 1. *Lipid Technology* 2:48–49.
- Copeman, L. A. and C. C. Parrish.** 2002. Lipid composition of malpigmented and normally pigmented newly settled yellowtail flounder, *Limanda ferruginea* (Storer). *Aquaculture Research* 33:1209–1219.
- Cuadros-Rodríguez, L., M. G. Bagur-González, M. Sánchez-Viñas, A. González-Casado, and A. M. Gómez-Sáez.** 2007. Principles of analytical calibration/quantification for the separation sciences. *Journal of Chromatography A* 1158:33–46.
- De Francesco, M., G. Parisi, J. Pérez-Sánchez, P. Gómez-Réqueni, F. Médale, S. J. Kaushik, M. Mecatti, and B. M. Poli.** 2007. Effect of high-level fish meal replacement by plant proteins in gilthead sea bream (*Sparus aurata*) on growth and body/fillet quality traits. *Aquaculture Nutrition* 13:361–372.
- De Silva, S. S., R. M. Gunasekera, and B. A. Ingram.** 2004. Performance of intensively farmed Murray cod *Maccullochella peelii peilii* (Mitchell) fed newly formulated vs. currently used commercial diets, and a comparison of fillet composition of farmed and wild fish. *Aquaculture Research* 35:1039–1052.
- Díaz-López, M., M. J. Pérez, N. G. Acosta, D. R. Tocher, S. Jerez, A. Lorenzo, and C. Rodríguez.** 2009. Effect of dietary substitution of fish oil by *Echium* oil on growth, plasma parameters and body lipid composition in gilthead seabream (*Sparus aurata* L.). *Aquaculture Nutrition* 15:500–512.
- Dodds, E. D., M. R. McCoy, L. D. Rea, and J. M. Kennish.** 2005. Gas chromatographic quantification of fatty acid methyl esters: flame ionization detection vs. electron impact mass spectrometry. *Lipids* 40:419–428.
- Dyerberg, J., H. O. Bang, and N. Hjerne.** 1975. Fatty acid composition of plasma lipids in Greenland Eskimos. *The American Journal of Clinical Nutrition* 28:958–966.
- Enser, M., K. G. Hallet, B. Hewett, G. A. J. Fursey, J. D. Wood, and G. Harrington.** 1998. Fatty acid content and composition of UK beef and lamb muscle in relation to production system and implications for human nutrition. *Meat Science* 49:329–341.
- Figueiredo-Silva, A. C., G. Corraze, P. Borges, and L. M. P. Valente.** 2010. Dietary protein/lipid level and protein source effects on growth, tissue composition and lipid metabolism of blackspot seabream (*Pagellus bogaraveo*). *Aquaculture Nutrition* 16:173–187.
- Folch, J., M. Lees, and G. H. Sloane Stanley.** 1957. A simple method for the isolation and purification of total lipids from animal tissues. *The Journal of Biological Chemistry* 226:497–509.
- García, A. S., C. C. Parrish, J. A. Brown, S. C. Johnson, and S. Leadbeater.** 2008. Use of differently enriched rotifers, *Brachionus plicatilis*, during larviculture of haddock, *Melanogrammus aeglefinus*: effects on early growth, survival and body lipid composition. *Aquaculture Nutrition* 14:431–444.
- Garduño-Lugo, M., J. R. Herrera-Solís, J. O. Angulo-Guerrero, G. Muñoz-Córdova, and J. De la Cruz-Medina.** 2007. Nutrient composition and sensory evaluation of fillets from wild-type Nile tilapia (*Oreochromis niloticus*, Linnaeus) and a red hybrid (Florida red tilapia x red *O. niloticus*). *Aquaculture Research* 38:1074–1081.
- Givens, D. I. and R. A. Gibbs.** 2008. Current intakes of EPA and DHA in European populations and the potential of animal-derived foods to increase them. *Proceedings of the Nutrition Society* 67:273–280.
- Hardy, R. W.** 2003. Farmed fish and omega-3 fatty acids. *Aquaculture Magazine* 29:63–65.
- Jankowska, B., Z. Zakęs, T. Żmijewski, and M. Szczepkowski.** 2008. Fatty acid composition of wild and cultured northern pike (*Esox lucius*). *Journal of Applied Ichthyology* 24:196–201.
- Kandemir, S. and N. Polat.** 2007. Seasonal variation of total lipid and total fatty acid in muscle and liver of rainbow trout (*Oncorhynchus mykiss* W., 1792) reared in Derbent Dam Lake. *Turkish Journal of Fisheries and Aquatic Sciences* 7:27–31.
- Kucska, B., L. Pál, T. Müller, M. Bódis, Á. Bartos, L. Wágner, F. Husvéth, and M. Bercsényi.** 2006. Changing of fat content and fatty acid profile of reared pike (*Esox lucius*) fed two different diets. *Aquaculture Research* 37:96–101.
- Ludwig, G. M., S. D. Rawles, and S. E. Lochmann.** 2008. Effect of rotifer enrichment on sunshine bass *Morone chrysops* x *M. saxatilis* larvae growth and survival and fatty acid composition. *Journal of the World Aquaculture Society* 39:158–173.
- Lund, I., S. J. Steinfeldt, K. I. Suhr, and B. W. Hansen.** 2008. A comparison of fatty acid composition and quality aspects of eggs and larvae from cultured and wild broodstock of common sole (*Solea solea* L.). *Aquaculture Nutrition* 14:544–555.
- Nguyen, V. T., S. Satoh, Y. Haga, H. Fushimi, and T. Kotani.** 2008. Effect of zinc and manganese supplementation in *Artemia* on growth and vertebral deformity in red sea bream (*Pagrus major*) larvae. *Aquaculture* 285:184–192.

- Olsson, G. B., R. L. Olsen, M. Carlehög, and R. Ofstad. 2003. Seasonal variations in chemical and sensory characteristics of farmed and wild Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture* 217: 191–205.
- Palmegiano, G. B., F. Daprá, G. Forneris, F. Gai, L. Gasco, K. Guo, P. G. Peiretti, B. Sicuro, and I. Zoccarato. 2006. Rice protein concentrate meal as a potential ingredient in practical diets for rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 258:257–267.
- Park, H. G., V. Puvanendran, A. Kellett, C. C. Parrish, and J. A. Brown. 2006. Effect of enriched rotifers on growth, survival, and composition of larval Atlantic cod (*Gadus morhua*). *ICES Journal of Marine Science* 63:285–295.
- Ramos, A., N. M. Bandarra, P. Rema, P. Vaz-Pires, M. L. Nunes, A. M. Andrade, A. R. Cordeiro, and L. M. P. Valente. 2008. Time course deposition of conjugated linoleic acid in market size rainbow trout (*Oncorhynchus mykiss*) muscle. *Aquaculture* 274:366–374.
- Robin, J. H. and A. Peron. 2004. Consumption vs. deposition of essential fatty acids in gilthead sea bream (*Sparus aurata*) larvae fed semi-purified diets. *Aquaculture* 238:283–294.
- Rosenlund, G., A. Obach, M. G. Sandberg, H. Standal, and K. Tveit. 2001. Effect of alternative lipid sources on long-term growth performance and quality of Atlantic salmon (*Salmo salar* L.). *Aquaculture Research* 32(Suppl. 1):323–328.
- Ruxton, C. H. S. and E. Derbyshire. 2009. Latest evidence on omega-3 fatty acids and health. *Nutrition and Food Science* 39:423–438.
- Schlechtriem, C., J. E. Bron, and D. R. Tocher. 2007. Inter-individual variation in total fatty acid compositions of flesh of Atlantic salmon smolts-fed diets containing fish oil or vegetable oil. *Aquaculture Research* 38:1045–1055.
- Schlechtriem, C., J. E. Bron, and D. R. Tocher. 2009. Determination of n-3 HUFA content in Atlantic salmon flesh based on the lipid content, morphometric measurements and blood fatty acid composition: a modelling approach. *Journal of Applied Ichthyology* 25:120–123.
- Schwertner, H. A. and E. L. Mosser. 1993. Comparison of lipid fatty acids on a concentration basis vs. weight percentage basis in patients with and without coronary artery disease or diabetes. *Clinical Chemistry* 39:659–663.
- Seychelles, L. H., C. Audet, R. Tremblay, K. Lemarchand, and F. Pernet. 2011. Bacterial colonization of winter flounder *Pseudopleuronectes americanus* fed live feed enriched with three different commercial diets. *Aquaculture Nutrition* 17:e196–e206.
- Sharma, P., V. Kumar, A. K. Sinha, J. Ranjan, H. M. P. Kithsiri, and G. Venkateshwarlu. 2010. Comparative fatty acid profiles of wild and farmed tropical freshwater fish rohu (*Labeo rohita*). *Fish Physiology and Biochemistry* 36:411–417.
- St-Hilaire, S., C. Sheppard, J. K. Tomberlin, S. Irving, L. Newton, M. A. McGuire, E. E. Mosley, R. W. Hardy, and W. Sealey. 2007. Fly prepupae as a feedstuff for rainbow trout, *Oncorhynchus mykiss*. *Journal of the World Aquaculture Society* 38:59–67.
- Tidwell, J. H., S. Coyle, and L. A. Bright. 2007. Effects of different types of dietary lipids on growth and fatty acid composition of largemouth bass. *North American Journal of Aquaculture* 69:257–264.
- Tocher, D. R. 2003. Metabolism and functions of lipids and fatty acids in teleost fish. *Reviews in Fisheries Science* 11:107–184.
- Turchini, G. M., D. S. Francis, and S. S. De Silva. 2006. Modification of tissue fatty acid composition in Murray cod *Maccullochella peelii peelii* (Mitchell) resulting from a shift from vegetable oil diets to a fish oil diet. *Aquaculture Research* 37:570–585.
- Turchini, G. M., D. S. Francis, and S. S. De Silva. 2007. A whole body, in vivo, fatty acid balance method to quantify PUFA metabolism (desaturation, elongation and beta-oxidation). *Lipids* 42:1065–1071.
- Villalta, M., A. Estévez, M. P. Bransden, and J. G. Bell. 2008. Arachidonic acid, arachidonic/eicosapentaenoic acid ratio, stearidonic acid and eicosanoids are involved in dietary-induced albinism in Senegal sole (*Solea senegalensis*). *Aquaculture Nutrition* 14: 120–128.
- Yildirim-Aksoy, M., R. Shelby, C. Lim, and P. H. Klesius. 2007. Growth performance and proximate and fatty acid compositions of channel catfish, *Ictalurus punctatus*, fed for different duration with a commercial diet supplemented with various levels of menhaden fish oil. *Journal of the World Aquaculture Society* 38:461–474.
- Zhu, P., C. C. Parrish, and J. A. Brown. 2003. Lipid and amino acid metabolism during early development of Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture International* 11:43–52.

Appendix: List of Papers Included in the Overview (n = 394).

Papers with publication year “2011” were available online up to August 2010

- Abi-Ayad, S.-M. E.-A., P. Kestemont, and C. Mélard. 2000. Dynamics of total lipids and fatty acids during embryogenesis and larval development of Eurasian perch (*Perca fluviatilis*). *Fish Physiology and Biochemistry* 23:233–243.
- Abi-Ayad, S.-M. E.-A., Z. Boutiba, C. Mélard, and P. Kestemont. 2004. Dynamics of total body fatty acids during early ontogeny of pikeperch (*Sander lucioperca*) larvae. *Fish Physiology and Biochemistry* 30:129–136.
- Agius, R. V., T. Watanabe, S. Satoh, V. Kiron, H. Imaizumi, T. Yamazaki, and K. Kawano. 2001.

- Supplementation of paprika as a carotenoid source in soft-dry pellets for broodstock yellowtail *Seriola quinqueradiata* (Temminck & Schlegel). *Aquaculture Research* 32:263–272.
- Ai, Q., K. Mai, B. Tan, W. Xu, Q. Duan, H. Ma, and L. Zhang.** 2006. Replacement of fish meal by meat and bone meal in diets for large yellow croaker, *Pseudosciaena crocea*. *Aquaculture* 260:255–263.
- Ai, Q. H., J. Z. Zhao, K. S. Mai, W. Xu, B. P. Tan, H. M. Ma, and Z. G. Liufu.** 2008. Optimal dietary lipid level for large yellow croaker (*Pseudosciaena crocea*) larvae. *Aquaculture Nutrition* 14:515–522.
- Alimuddin, V. Kiron, S. Satoh, T. Takeuchi, and G. Yoshizaki.** 2008. Cloning and over-expression of a masu salmon (*Oncorhynchus masou*) fatty acid elongase-like gene in zebrafish. *Aquaculture* 282:13–18.
- Aliyu-Paiko, M., R. Hashim, A. Shu-Chien Chong, L. Yogarajah, and A.-F. M. El-Sayed.** 2010. Influence of different sources and levels of dietary protein and lipid on the growth, feed efficiency, muscle composition and fatty acid profile of Snakehead *Channa striatus* (Bloch, 1793) fingerling. *Aquaculture Research* 41:1365–1376.
- Almansa, E., M. V. Martían, J. R. Cejas, P. Badía, S. Jerez, and A. Lorenzo.** 2001. Lipid and fatty acid composition of female gilthead seabream during their reproductive cycle: effects of a diet lacking n-3 HUFA. *Journal of Fish Biology* 59:267–286.
- Arslan, M., J. Rinchar, K. Dabrowski, and M. C. Portella.** 2008. Effects of different dietary lipid sources on the survival, growth, and fatty acid composition of South American catfish, *Pseudoplatystoma fasciatum*, surubim, juveniles. *Journal of the World Aquaculture Society* 39:51–61.
- Arslan, M., K. Dabrowski, and M. C. Portella.** 2009. Growth, fat content and fatty acid profile of South American catfish, surubim (*Pseudoplatystoma fasciatum*) juveniles fed live, commercial and formulated diets. *Journal of Applied Ichthyology* 25:73–78.
- Asdari, R., M. Aliyu-Paiko, R. Hashim, and S. Ramachandran.** 2011. Effects of different dietary lipid sources in the diet for *Pangasius hypophthalmus* (Sauvage, 1878) juvenile on growth performance, nutrient utilization, body indices and muscle and liver fatty acid composition. *Aquaculture Nutrition* 17:44–53.
- Atalah, E., C. M. Hernández-Cruz, M. S. Izquierdo, G. Rosenlund, M. J. Caballero, A. Valencia, and L. Robaina.** 2007. Two microalgae *Cryptocodinium cohnii* and *Phaeodactylum tricornutum* as alternative source of essential fatty acids in starter feeds for seabream (*Sparus aurata*). *Aquaculture* 270:178–185.
- Atwood, H. L., J. R. Tomasso, K. Webb, and D. M. Gatlin.** 2003. Low-temperature tolerance of Nile tilapia, *Oreochromis niloticus*: effects of environmental and dietary factors. *Aquaculture Research* 34:241–251.
- Bahurmiz, O. M. and W.-K. Ng.** 2007. Effects of dietary palm oil source on growth, tissue fatty acid composition and nutrient digestibility of red hybrid tilapia, *Oreochromis* sp., raised from stocking to marketable size. *Aquaculture* 262:382–392.
- Bake, G. G., M. Endo, A. Akimoto, and T. Takeuchi.** 2009. Evaluation of recycled food waste as a partial replacement of fishmeal in diets for the initial feeding of Nile tilapia *Oreochromis niloticus*. *Fisheries Science* 75:1275–1283.
- Bandarra, N. M., M. L. Nunes, A. M. Andrade, J. A. M. Prates, S. Pereira, M. Monteiro, P. Rema, and L. M. P. Valente.** 2006. Effect of dietary conjugated linoleic acid on muscle, liver and visceral lipid deposition in rainbow trout juveniles (*Oncorhynchus mykiss*). *Aquaculture* 254:496–505.
- Baskerville-Bridges, B. and L. J. Kling.** 2000. Development and evaluation of microparticulate diets for early weaning of Atlantic cod *Gadus morhua* larvae. *Aquaculture Nutrition* 6:171–182.
- Battaglene, S. C., D. T. Morehead, J. M. Cobcroft, P. D. Nichols, M. R. Brown, and J. Carson.** 2006. Combined effects of feeding enriched rotifers and antibiotic addition on performance of striped trumpeter (*Latris lineata*) larvae. *Aquaculture* 251:456–471.
- Bell, M. V. and J. R. Dick.** 2004. Changes in capacity to synthesise 22:6n-3 during early development in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 235:393–409.
- Bell, J. G., F. McGhee, P. J. Campbell, and J. R. Sargent.** 2003. Rapeseed oil as an alternative to marine fish oil in diets of post-smolt Atlantic salmon (*Salmo salar*): changes in flesh fatty acid composition and effectiveness of subsequent fish oil “wash out.” *Aquaculture* 218:515–528.
- Bell, J. G., F. McGhee, J. R. Dick, and D. R. Tocher.** 2005. Dioxin and dioxin-like polychlorinated biphenyls (PCBs) in Scottish farmed salmon (*Salmo salar*): effects of replacement of dietary marine fish oil with vegetable oils. *Aquaculture* 243:305–314.
- Bell, J. G., F. Strachan, J. E. Good, and D. R. Tocher.** 2006. Effect of dietary echium oil on growth, fatty acid composition and metabolism, gill prostaglandin production and macrophage activity in Atlantic cod (*Gadus morhua* L.). *Aquaculture Research* 37:606–617.
- Bell, J. G., J. Pratoomyot, F. Strachan, R. J. Henderson, R. Fontanillas, A. Hebard, D. R. Guy, D. Hunter, and D. R. Tocher.** 2010. Growth, flesh adiposity and fatty acid composition of Atlantic salmon (*Salmo salar*) families with contrasting flesh adiposity: effects of replacement of dietary fish oil with vegetable oils. *Aquaculture* 306:225–232.
- Bendiksen, E. Å., A. M. Arnesen, and M. Jobling.** 2003. Effects of dietary fatty acid profile and fat content on smolting and seawater performance in Atlantic salmon (*Salmo salar* L.). *Aquaculture* 225:149–163.
- Benedito-Palos, L., J. C. Navarro, A. Bermejo-Nogales, A. Saera-Vila, S. Kaushik, and J. Pérez-Sánchez.**

2009. The time course of fish oil wash-out follows a simple dilution model in gilthead sea bream (*Sparus aurata* L.) fed graded levels of vegetable oils. *Aquaculture* 288:98–105.
- Berge, G. M., B. Ruyter, and T. Åsgård. 2004. Conjugated linoleic acid in diets for juvenile Atlantic salmon (*Salmo salar*); effects on fish performance, proximate composition, fatty acid and mineral content. *Aquaculture* 237:365–380.
- Berntssen, M. H. G., A.-K. Lundebye, and B. E. Torstensen. 2005. Reducing the levels of dioxins and dioxin-like PCBs in farmed Atlantic salmon by substitution of fish oil with vegetable oil in the feed. *Aquaculture Nutrition* 11:219–231.
- Betancor, M. B., E. Atalah, M. Caballero, T. Benítez-Santana, J. Roo, D. Montero, and M. Izquierdo. 2011. α -Tocopherol in weaning diets for European sea bass (*Dicentrarchus labrax*) improves survival and reduces tissue damage caused by excess dietary DHA contents. *Aquaculture Nutrition* 17:e112–e122.
- Biswas, A. K., J. Nozaki, M. Kurata, K. Takii, H. Kumai, and M. Seoka. 2006. Effect of *Artemia* enrichment on the growth and survival of Pacific bluefin tuna *Thunnus orientalis* (Temminck & Schlegel) larvae. *Aquaculture Research* 37:1662–1670.
- Biswas, B. K., S.-C. Ji, A. K. Biswas, M. Seoka, Y.-S. Kim, K.-I. Kawasaki, and K. Takii. 2009. Dietary protein and lipid requirements for the Pacific bluefin tuna *Thunnus orientalis* juvenile. *Aquaculture* 288:114–119.
- Blair, T., J. Castell, S. Neil, L. D'Abramo, C. Cahu, P. Harmon, and K. Ogunmoye. 2003. Evaluation of microdiets versus live feeds on growth, survival and fatty acid composition of larval haddock (*Melanogrammus aeglefinus*). *Aquaculture* 225:451–461.
- Blanchard, G., J. G. Makombu, and P. Kestemont. 2008. Influence of different dietary 18:3n-3/18:2n-6 ratio on growth performance, fatty acid composition and hepatic ultrastructure in Eurasian perch, *Perca fluviatilis*. *Aquaculture* 284:144–150.
- Bogut, I., E. Has-Schön, M. Čačić, Z. Milaković, D. Novoselić, and S. Brkić. 2002. Linolenic acid supplementation in the diet of European catfish (*Silurus glanis*): effect on growth and fatty acid composition. *Journal of Applied Ichthyology* 18:1–6.
- Brandsen, M. P., J. M. Cobcroft, S. C. Battaglene, G. A. Dunstan, P. D. Nichols, and J. G. Bell. 2004. Dietary arachidonic acid alters tissue fatty acid profile, whole body eicosanoid production and resistance to hypersaline challenge in larvae of the temperate marine fish, striped trumpeter (*Latris lineata*). *Fish Physiology and Biochemistry* 30:241–256.
- Brandsen, M. P., G. M. Butterfield, J. Walden, L. A. McEvoy, and J. G. Bell. 2005a. Tank colour and dietary arachidonic acid affects pigmentation, eicosanoid production and tissue fatty acid profile of larval Atlantic cod (*Gadus morhua*). *Aquaculture* 250:328–340.
- Brandsen, M. P., S. C. Battaglene, D. T. Morehead, G. A. Dunstan, and P. D. Nichols. 2005b. Effect of dietary 22:6n-3 on growth, survival and tissue fatty acid profile of striped trumpeter (*Latris lineata*) larvae fed enriched *Artemia*. *Aquaculture* 243:331–344.
- Brown, M. R., G. A. Dunstan, P. D. Nichols, S. C. Battaglene, D. T. Morehead, and A. L. Overweter. 2005. Effects of [alpha]-tocopherol supplementation of rotifers on the growth of striped trumpeter *Latris lineata* larvae. *Aquaculture* 246:367–378.
- Brown, T. D., D. S. Francis, and G. M. Turchini. 2010. Can dietary lipid source circadian alternation improve omega-3 deposition in rainbow trout? *Aquaculture* 300:148–155.
- Bureau, D. P., K. Hua, and A. M. Harris. 2008. The effect of dietary lipid and long-chain n-3 PUFA levels on growth, energy utilization, carcass quality, and immune function of rainbow trout, *Oncorhynchus mykiss*. *Journal of the World Aquaculture Society* 39:1–21.
- Busch, K. E. T., I.-B. Falk-Petersen, S. Peruzzi, N. A. Rist, and K. Hamre. 2010. Natural zooplankton as larval feed in intensive rearing systems for juvenile production of Atlantic cod (*Gadus morhua* L.). *Aquaculture Research* 41:1727–1740.
- Caballero, M. J., A. Obach, G. Rosenlund, D. Montero, M. Gisvold, and M. S. Izquierdo. 2002. Impact of different dietary lipid sources on growth, lipid digestibility, tissue fatty acid composition and histology of rainbow trout, *Oncorhynchus mykiss*. *Aquaculture* 214:253–271.
- Castell, J., T. Blair, S. Neil, K. Howes, S. Mercer, J. Reid, W. Young-Lai, B. Gullison, P. Dhert, and P. Sorgeloos. 2003. The effect of different HUFA enrichment emulsions on the nutritional value of rotifers (*Brachionus plicatilis*) fed to larval haddock (*Melanogrammus aeglefinus*). *Aquaculture International* 11:109–117.
- Cavalin, F. G. and C. R. Weirich. 2009. Larval performance of aquacultured Florida pompano (*Trachinotus carolinus*) fed rotifers (*Brachionus plicatilis*) enriched with selected commercial diets. *Aquaculture* 292:67–73.
- Cejas, J. R., E. Almansa, N. Tejera, S. Jerez, A. Bolaños, and A. Lorenzo. 2003. Effect of dietary supplementation with shrimp on skin pigmentation and lipid composition of red porgy (*Pagrus pagrus*) alevins. *Aquaculture* 218:457–469.
- Chan, J. C. K., J. Mann, B. J. Skura, M. Rowshandeli, N. Rowshandeli, and D. A. Higgs. 2002. Effects of feeding diets containing various dietary protein and lipid ratios on the growth performance and pigmentation of post-juvenile coho salmon *Oncorhynchus kisutch* reared in sea water. *Aquaculture Research* 33:1137–1156.
- Chatzifotis, S., P. Muje, M. Pavlidis, J. Ågren, M. Paalavuo, and H. Mölsä. 2004. Evolution of tissue composition and serum metabolites during gonadal

- development in the common dentex (*Dentex dentex*). *Aquaculture* 236:557–573.
- Cho, S. H., S. M. Lee, S. M. Lee, and J. H. Lee.** 2005. Effect of dietary protein and lipid levels on growth and body composition of juvenile turbot (*Scophthalmus maximus* L.) reared under optimum salinity and temperature conditions. *Aquaculture Nutrition* 11:235–240.
- Chou, B.-S., S.-Y. Shiau, and S. S. O. Hung.** 2001. Effect of dietary cod liver oil on growth and fatty acids of juvenile hybrid tilapia. *North American Journal of Aquaculture* 63:277–284.
- Civera-Cerecedo, R., C. A. Alvarez-González, R. E. García-Gómez, V. Carrasco-Chávez, J. L. Ortiz-Galindo, M. O. Rosales-Velázquez, T. Grayeb-Del Álamo, and F. J. Moyano-López.** 2008. Effect of microparticulate diets on growth and survival of spotted sand bass larvae, *Paralabrax maculatofasciatus*, at two early weaning times. *Journal of the World Aquaculture Society* 39:22–36.
- Copeman, L. A. and C. C. Parrish.** 2002. Lipid composition of malpigmented and normally pigmented newly settled yellowtail flounder, *Limanda ferruginea* (Storer). *Aquaculture Research* 33:1209–1219.
- Copeman, L. A., C. C. Parrish, J. A. Brown, and M. Harel.** 2002. Effects of docosahexaenoic, eicosapentaenoic, and arachidonic acids on the early growth, survival, lipid composition and pigmentation of yellowtail flounder (*Limanda ferruginea*): a live food enrichment experiment. *Aquaculture* 210:285–304.
- Crampton, V. O., D. A. Nanton, K. Ruohonen, P. O. Skjervold, and A. El-Mowafi.** 2010. Demonstration of salmon farming as a net producer of fish protein and oil. *Aquaculture Nutrition* 16:437–446.
- Cutts, C. J., J. Sawanboonchun, C. Mazorra de Quero, and J. G. Bell.** 2006. Diet-induced differences in the essential fatty acid (EFA) compositions of larval Atlantic cod (*Gadus morhua* L.) with reference to possible effects of dietary EFAs on larval performance. *ICES Journal of Marine Science* 63:302–310.
- D'Abramo, L. R., C. L. Ohs, and J. B. Taylor.** 2000. Effects of reduced levels of dietary protein and menhaden fish meal on production, dressout, and biochemical composition of phase III sunshine bass *Morone chrysops* ♀ × *M. saxatilis* ♂ cultured in earthen ponds. *Journal of the World Aquaculture Society* 31:316–325.
- Dâmaso-Rodrigues, M. L., P. Pousão-Ferreira, L. Ribeiro, J. Coutinho, N. M. Bandarra, P. J. Gavaia, L. Narciso, and S. Morais.** 2010. Lack of essential fatty acids in live feed during larval and post-larval rearing: effect on the performance of juvenile *Solea senegalensis*. *Aquaculture International* 18:741–757.
- Dantagnan, P., A. Hernández, A. Borquez, and A. Mansilla.** 2009. Inclusion of macroalgae meal (*Macrocystis pyrifera*) as feed ingredient for rainbow trout (*Oncorhynchus mykiss*): effect on flesh fatty acid composition. *Aquaculture Research* 41:87–94.
- De Francesco, M., G. Parisi, F. Médale, P. Lupi, S. J. Kaushik, and B. M. Poli.** 2004. Effect of long-term feeding with a plant protein mixture based diet on growth and body/fillet quality traits of large rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 236:413–429.
- De Francesco, M., G. Parisi, J. Pérez-Sánchez, P. Gómez-Réqueni, F. Médale, S. J. Kaushik, M. Mecatti, and B. M. Poli.** 2007. Effect of high-level fish meal replacement by plant proteins in gilthead sea bream (*Sparus aurata*) on growth and body/fillet quality traits. *Aquaculture Nutrition* 13:361–372.
- De la Pena, M. R.** 2001. Use of juvenile instar *Diaphanosoma calebensis* (Stingelin) in hatchery rearing of Asian sea bass *Lates calcarifer* (Bloch). *The Israeli Journal of Aquaculture* 53:128–138.
- De Silva, S. S., R. M. Gunasekera, R. A. Collins, and B. A. Ingram.** 2002. Performance of juvenile Murray cod, *Maccullochella peelii peelii* (Mitchell), fed with diets of different protein to energy ratio. *Aquaculture Nutrition* 8:79–85.
- De Silva, S. S., R. M. Gunasekera, and B. A. Ingram.** 2004. Performance of intensively farmed Murray cod *Maccullochella peelii peelii* (Mitchell) fed newly formulated vs. currently used commercial diets, and a comparison of fillet composition of farmed and wild fish. *Aquaculture Research* 35:1039–1052.
- Díaz-López, M., M. J. Pérez, N. G. Acosta, D. R. Tocher, S. Jerez, A. Lorenzo, and C. Rodríguez.** 2009. Effect of dietary substitution of fish oil by *Echium* oil on growth, plasma parameters and body lipid composition in gilthead seabream (*Sparus aurata* L.). *Aquaculture Nutrition* 15:500–512.
- Domaizon, I., C. Desvillettes, D. Debroas, and G. Bourdier.** 2000. Influence of zooplankton and phytoplankton on the fatty acid composition of digesta and tissue lipids of silver carp: mesocosm experiment. *Journal of Fish Biology* 57:417–432.
- Dos Santos, L. D., W. M. Furuya, L. C. R. Da Silva, M. Matsushita, and T. S. De Castro Silva.** 2011. Dietary conjugated linoleic acid (CLA) for finishing Nile tilapia. *Aquaculture Nutrition* 17:e70–e81.
- Drew, M. D., A. E. Ogunkoya, D. M. Janz, and A. G. Van Kessel.** 2007. Dietary influence of replacing fish meal and oil with canola protein concentrate and vegetable oils on growth performance, fatty acid composition and organochlorine residues in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 267:260–268.
- Du, Z. Y., P. Clouet, L. M. Huang, P. Degrace, W. H. Zheng, J. G. He, L. X. Tian, and Y. J. Liu.** 2008. Utilization of different dietary lipid sources at high level in herbivorous grass carp (*Ctenopharyngodon idella*): mechanism related to hepatic fatty acid oxidation. *Aquaculture Nutrition* 14:77–92.
- El-Husseiny, O. M., G. M. Abdul-Aziz, A. M. A.-S. Goda, and A. Suloma.** 2010. Effect of altering linoleic acid and linolenic acid dietary levels and ratios on the performance and tissue fatty acid profiles of

- Nile tilapia *Oreochromis niloticus* fry. *Aquaculture International* 18:1105–1119.
- Eusebio, P. S., R. M. Coloso, and R. S. J. Gapasin.** 2010. Nutritional evaluation of mysids *Mesopodopsis orientalis* (Crustacea: Mysida) as live food for grouper *Epinephelus fuscoguttatus* larvae. *Aquaculture* 306:289–294.
- Evjemo, J. O., K. I. Reitan, and Y. Olsen.** 2003. Copepods as live food organisms in the larval rearing of halibut larvae (*Hippoglossus hippoglossus* L.) with special emphasis on the nutritional value. *Aquaculture* 227:191–210.
- Faleiro, F. and L. Narciso.** 2010. Lipid dynamics during early development of *Hippocampus guttulatus* seahorses: Searching for clues on fatty acid requirements. *Aquaculture* 307:56–64.
- Faulk, C. K. and G. J. Holt.** 2003. Lipid nutrition and feeding of cobia *Rachycentron canadum* larvae. *Journal of the World Aquaculture Society* 34:368–378.
- Faulk, C. K. and G. J. Holt.** 2005. Advances in rearing cobia *Rachycentron canadum* larvae in recirculating aquaculture systems: live prey enrichment and green-water culture. *Aquaculture* 249:231–243.
- Faulk, C. K., G. J. Holt, and D. A. Davis.** 2005. Evaluation of fatty acid enrichment of live food for yellowtail snapper *Ocyurus chrysurus* larvae. *Journal of the World Aquaculture Society* 36:271–281.
- Figueiredo-Silva, A. C., G. Corraze, P. Rema, J. Sanchez-Gurmaches, J. Gutiérrez, and L. M. P. Valente.** 2009. Blackspot seabream (*Pagellus bogaraveo*) lipogenic and glycolytic pathways appear to be more related to dietary protein level than dietary starch type. *Aquaculture* 291:101–110.
- Figueiredo-Silva, A. C., G. Corraze, P. Borges, and L. M. P. Valente.** 2010. Dietary protein/lipid level and protein source effects on growth, tissue composition and lipid metabolism of blackspot seabream (*Pagellus bogaraveo*). *Aquaculture Nutrition* 16:173–187.
- Fonseca-Madrugal, J., V. Karalazos, P. J. Campbell, J. G. Bell, and D. R. Tocher.** 2005. Influence of dietary palm oil on growth, tissue fatty acid compositions, and fatty acid metabolism in liver and intestine in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture Nutrition* 11:241–250.
- Fontagné, S., G. Corraze, and P. Bergot.** 2000a. Response of common carp (*Cyprinus carpio*) larvae to different dietary levels and forms of supply of medium-chain fatty acids. *Aquatic Living Resources* 13:429–437.
- Fontagné, S., J. Robin, G. Corraze, and P. Bergot.** 2000b. Growth and survival of European sea bass (*Dicentrarchus labrax*) larvae fed from first feeding on compound diets containing medium-chain triacylglycerols. *Aquaculture* 190:261–271.
- Fontagné, S., L. Burtaire, G. Corraze, and P. Bergot.** 2000c. Effects of dietary medium-chain triacylglycerols (tricaprylin and tricaproin) and phospholipid supply on survival, growth and lipid metabolism in common carp (*Cyprinus carpio* L.) larvae. *Aquaculture* 190:289–303.
- Fountoulaki, E., M. N. Alexis, I. Nengas, and B. Venou.** 2003. Effects of dietary arachidonic acid (20:4n-6), on growth, body composition, and tissue fatty acid profile of gilthead bream fingerlings (*Sparus aurata* L.). *Aquaculture* 225:309–323.
- Fountoulaki, E., A. Vasilaki, R. Hurtado, K. Grigorakis, I. Karacostas, I. Nengas, G. Rigos, Y. Kotzamanis, B. Venou, and M. N. Alexis.** 2009. Fish oil substitution by vegetable oils in commercial diets for gilthead sea bream (*Sparus aurata* L.); effects on growth performance, flesh quality and fillet fatty acid profile: Recovery of fatty acid profiles by a fish oil finishing diet under fluctuating water temperatures. *Aquaculture* 289:317–326.
- Francis, D. S., G. M. Turchini, P. L. Jones, and S. S. De Silva.** 2006. Effects of dietary oil source on growth and fillet fatty acid composition of Murray cod, *Maccullochella peelii peelii*. *Aquaculture* 253:547–556.
- Francis, D. S., G. M. Turchini, P. L. Jones, and S. S. De Silva.** 2007. Growth performance, feed efficiency and fatty acid composition of juvenile Murray cod, *Maccullochella peelii peelii*, fed graded levels of canola and linseed oil. *Aquaculture Nutrition* 13:335–350.
- Francis, D. S., G. M. Turchini, B. K. Smith, S. G. Ryan, and S. S. De Silva.** 2009. Effects of alternate phases of fish oil and vegetable oil-based diets in Murray cod. *Aquaculture Research* 40:1123–1134.
- Ganuza, E., T. Benítez-Santana, E. Atalah, O. Vega-Orellana, R. Ganga, and M. S. Izquierdo.** 2008. *Cryptocodinium cohnii* and *Schizochytrium* sp. as potential substitutes to fisheries-derived oils from seabream (*Sparus aurata*) microdiets. *Aquaculture* 277:109–116.
- Gapasin, R. S. J. and M. N. Duray.** 2001. Effects of DHA-enriched live food on growth, survival and incidence of opercular deformities in milkfish (*Chanos chanos*). *Aquaculture* 193:49–63.
- García de La Banda, I., C. Lobo, J. M. León-Rubio, S. Tapia-Paniagua, M. C. Balebona, M. A. Morínigo, X. Moreno-Ventas, L. M. Lucas, F. Linares, F. Arce, and S. Arijo.** 2010. Influence of two closely related probiotics on juvenile Senegalese sole (*Solea senegalensis*, Kaup 1858) performance and protection against *Photobacterium damsela* subsp. *piscicida*. *Aquaculture* 306:281–288.
- Garcia, A. S., C. C. Parrish, and J. A. Brown.** 2008a. Growth and lipid composition of Atlantic cod (*Gadus morhua*) larvae in response to differently enriched *Artemia franciscana*. *Fish Physiology and Biochemistry* 34:77–94.
- Garcia, A. S., C. C. Parrish, and J. A. Brown.** 2008b. A comparison among differently enriched rotifers (*Brachionus plicatilis*) and their effect on Atlantic cod (*Gadus morhua*) larvae early growth, survival and lipid composition. *Aquaculture Nutrition* 14:14–30.
- Garcia, A. S., C. C. Parrish, J. A. Brown, S. C. Johnson, and S. Leadbeater.** 2008c. Use of differently enriched rotifers, *Brachionus plicatilis*, during larviculture of

- haddock, *Melanogrammus aeglefinus*: effects on early growth, survival and body lipid composition. *Aquaculture Nutrition* 14:431–444.
- Gardeur, J.-N., N. Mathis, A. Kobilinsky, and J. Brun-Bellut.** 2007. Simultaneous effects of nutritional and environmental factors on growth and flesh quality of *Perca fluviatilis* using a fractional factorial design study. *Aquaculture* 273:50–63.
- Garduño-Lugo, M., J. R. Herrera-Solís, J. O. Angulo-Guerrero, G. Muñoz-Córdova, and J. De la Cruz-Medina.** 2007. Nutrient composition and sensory evaluation of fillets from wild-type Nile tilapia (*Oreochromis niloticus*, Linnaeus) and a red hybrid (Florida red tilapia × red *O. niloticus*). *Aquaculture Research* 38:1074–1081.
- Gasco, L., F. Gai, C. Lussiana, R. Lo Presti, V. Malfatto, F. Daprà, and I. Zoccarato.** 2010. Morphometry, slaughtering performances, chemical and fatty acid composition of the protected designation of origin “Golden hump tench of Poirino highland” product. *Reviews in Fish Biology and Fisheries* 20:357–365.
- Gawlicka, A., M. A. Herold, F. T. Barrows, J. De La Noüe, and S. S. O. Hung.** 2002. Effects of dietary lipids on growth, fatty acid composition, intestinal absorption and hepatic storage in white sturgeon (*Acipenser transmontanus* R.) larvae. *Journal of Applied Ichthyology* 18:673–681.
- Giménez, G., A. Estévez, R. J. Henderson, and J. G. Bell.** 2008. Changes in lipid content, fatty acid composition and lipid class composition of eggs and developing larvae (0–40 days old) of cultured common dentex (*Dentex dentex* Linnaeus 1758). *Aquaculture Nutrition* 14:300–308.
- Glencross, B., W. Hawkins, and J. Curnow.** 2003a. Evaluation of canola oils as alternative lipid resources in diets for juvenile red seabream, *Pagrus auratus*. *Aquaculture Nutrition* 9:305–315.
- Glencross, B. D., W. E. Hawkins, and J. G. Curnow.** 2003b. Restoration of the fatty acid composition of red seabream (*Pagrus auratus*) using a fish oil finishing diet after grow-out on plant oil based diets. *Aquaculture Nutrition* 9:409–418.
- Glover, K. A., H. Otterå, R. E. Olsen, E. Slinde, G. L. Taranger, and Ø. Skaala.** 2009. A comparison of farmed, wild and hybrid Atlantic salmon (*Salmo salar* L.) reared under farming conditions. *Aquaculture* 286:203–210.
- Grageda, M. V. C., T. Kotani, Y. Sakakura, and A. Hagiwara.** 2008. Effects of feeding copepod and Artemia on early growth and behaviour of the self-fertilizing fish, *Rivulus marmoratus*, under laboratory conditions. *Aquaculture* 281:100–105.
- Grant, A. A. M., D. Baker, D. A. Higgs, C. J. Brauner, J. G. Richards, S. K. Balfry, and P. M. Schulte.** 2008. Effects of dietary canola oil level on growth, fatty acid composition and osmoregulatory ability of juvenile fall chinook salmon (*Oncorhynchus tshawytscha*). *Aquaculture* 277:303–312.
- Grigorakis, K., E. Fountoulaki, I. Giogios, and M. N. Alexis.** 2009. Volatile compounds and organoleptic qualities of gilthead sea bream (*Sparus aurata*) fed commercial diets containing different lipid sources. *Aquaculture* 290:116–121.
- Grisdale-Helland, B., B. Ruyter, G. Rosenlund, A. Obach, S. J. Helland, M. G. Sandberg, H. Standal, and C. Røsjø.** 2002. Influence of high contents of dietary soybean oil on growth, feed utilization, tissue fatty acid composition, heart histology and standard oxygen consumption of Atlantic salmon (*Salmo salar*) raised at two temperatures. *Aquaculture* 207:311–329.
- Gunasekera, R. M., S. S. De Silva, and B. A. Ingram.** 2001. Chemical changes in fed and starved larval trout cod, *Maccullochella macquarensis* during early development. *Fish Physiology and Biochemistry* 25:255–268.
- Hafezieh, M., M. S. S. Kamarudin, C. R. Bin Saad, M. K. Abd Sattar, N. Agh, T. Valinassab, M. Sharifian, and H. Hosseinpour.** 2010. Effects of enriched *Artemia urmiana* with HUFA on growth, survival, and fatty acids composition of the Persian sturgeon larvae (*Acipenser persicus*). *Iranian Journal of Fisheries Sciences* 9:61–72.
- Hamre, K. and T. Harboe.** 2008a. Artemia enriched with high n-3 HUFA may give a large improvement in performance of Atlantic halibut (*Hippoglossus hippoglossus* L.) larvae. *Aquaculture* 277:239–243.
- Hamre, K. and T. Harboe.** 2008b. Critical levels of essential fatty acids for normal pigmentation in Atlantic halibut (*Hippoglossus hippoglossus* L.) larvae. *Aquaculture* 277:101–108.
- Hamre, K., I. Opstad, M. Espe, J. Solbakken, G. I. Hemre, and K. Pittman.** 2002. Nutrient composition and metamorphosis success of Atlantic halibut (*Hippoglossus hippoglossus*, L.) larvae fed natural zooplankton or *Artemia*. *Aquaculture Nutrition* 8:139–148.
- Hamre, K., T. A. Mollan, Ø. Sæle, and B. Erstad.** 2008. Rotifers enriched with iodine and selenium increase survival in Atlantic cod (*Gadus morhua*) larvae. *Aquaculture* 284:190–195.
- Hamza, N., M. Mhetli, I. B. Khemis, C. Cahu, and P. Kestemont.** 2008. Effect of dietary phospholipid levels on performance, enzyme activities and fatty acid composition of pikeperch (*Sander lucioperca*) larvae. *Aquaculture* 275:274–282.
- Hansen, J. Ø., G. M. Berge, M. Hillestad, Å. Krogdahl, T. F. Galloway, H. Holm, J. Holm, and B. Ruyter.** 2008. Apparent digestion and apparent retention of lipid and fatty acids in Atlantic cod (*Gadus morhua*) fed increasing dietary lipid levels. *Aquaculture* 284:159–166.
- Harboe, T., A. Mangor-Jensen, M. Moren, K. Hamre, and I. Rønnestad.** 2009. Control of light condition affects the feeding regime and enables successful eye migration in Atlantic halibut juveniles. *Aquaculture* 290:250–255.

- Harel, M., S. Gavasso, J. Leshin, A. Gubernatis, and A. R. Place. 2001. The effect of tissue docosahexaenoic and arachidonic acids levels on hypersaline tolerance and leucocyte composition in striped bass (*Morone saxatilis*) larvae. *Fish Physiology and Biochemistry* 24:113–123.
- Harel, M., W. Koven, I. Lein, Y. Bar, P. Behrens, J. Stubblefield, Y. Zohar, and A. R. Place. 2002. Advanced DHA, EPA and ArA enrichment materials for marine aquaculture using single cell heterotrophs. *Aquaculture* 213:347–362.
- Hasimoglu, A., A. Erteken, S. Kino, and H. Nakagawa. 2007. Evaluation of anchovy meal and soybean meal as dietary protein sources for the Black Sea turbot, *Psetta maxima*. *The Israeli Journal of Aquaculture* 59:73–80.
- Haugen, T., A. Kiessling, R. E. Olsen, M. B. Rørå, E. Slinde, and R. Nortvedt. 2006. Seasonal variations in muscle growth dynamics and selected quality attributes in Atlantic halibut (*Hippoglossus hippoglossus* L.) fed dietary lipids containing soybean and/or herring oil under different rearing regimes. *Aquaculture* 261:565–579.
- Hedrick, R. L., T. J. Popma, and D. A. Davis. 2005. Pond production and fatty acid profiles of the filets of channel catfish reared on diets with different protein sources. *North American Journal of Aquaculture* 67:304–311.
- Hemre, G.-I. and K. Sandnes. 2008. Seasonal adjusted diets to Atlantic salmon (*Salmo salar*): Evaluations of a novel feed based on heat-coagulated fish mince, fed throughout 1 year in sea: Feed utilisation, retention of nutrients and health parameters. *Aquaculture* 274:166–174.
- Hemre, G. I., M. Sanden, A. M. Bakke-Mckellep, A. Sagstad, and Å. Kroghdahl. 2005. Growth, feed utilization and health of Atlantic salmon *Salmo salar* L. fed genetically modified compared to non-modified commercial hybrid soybeans. *Aquaculture Nutrition* 11:157–167.
- Higgs, D. A., S. K. Balfry, J. D. Oakes, M. Rowshandeli, B. J. Skura, and G. Deacon. 2006. Efficacy of an equal blend of canola oil and poultry fat as an alternate dietary lipid source for Atlantic salmon (*Salmo salar* L.) in sea water. I: effects on growth performance, and whole body and fillet proximate and lipid composition. *Aquaculture Research* 37:180–191.
- Hosseini, S. V., A. A. Kenari, J. M. Regenstein, M. Rezaei, R. M. Nazari, M. Moghaddasi, S. A. Kaboli, and A. A. M. Grant. 2010. Effects of alternative dietary lipid sources on growth performance and fatty acid composition of Beluga sturgeon, *Huso huso*, juveniles. *Journal of the World Aquaculture Society* 41:471–489.
- Huang, C.-H., W.-J. Shyong, and W.-Y. Lin. 2001. Dietary lipid supplementation affects the body fatty acid composition but not the growth of juvenile river chub, *Zacco barbata* (Regan). *Aquaculture Research* 32:1005–1010.
- Huang, S. S. Y., A. N. Oo, D. A. Higgs, C. J. Brauner, and S. Satoh. 2007. Effect of dietary canola oil level on the growth performance and fatty acid composition of juvenile red sea bream, *Pagrus major*. *Aquaculture* 271:420–431.
- Huang, S. S. Y., C. H. L. Fu, D. A. Higgs, S. K. Balfry, P. M. Schulte, and C. J. Brauner. 2008. Effects of dietary canola oil level on growth performance, fatty acid composition and ionoregulatory development of spring chinook salmon parr, *Oncorhynchus tshawytscha*. *Aquaculture* 274:109–117.
- Hunt, A. Ö., F. Özkan, K. Engin, and N. Tekelioğlu. 2011. The effects of freshwater rearing on the whole body and muscle tissue fatty acid profile of the European sea bass (*Dicentrarchus labrax*). *Aquaculture International* 19:51–61.
- Ibarz, A., J. Blasco, M. Beltrán, M. A. Gallardo, J. Sánchez, R. Sala, and J. Fernández-Borràs. 2005. Cold-induced alterations on proximate composition and fatty acid profiles of several tissues in gilthead sea bream (*Sparus aurata*). *Aquaculture* 249:477–486.
- Ibarz, A., M. Beltrán, J. Fernández-Borràs, M. A. Gallardo, J. Sánchez, and J. Blasco. 2007. Alterations in lipid metabolism and use of energy depots of gilt-head sea bream (*Sparus aurata*) at low temperatures. *Aquaculture* 262:470–480.
- Ibeas, C., C. Rodríguez, P. Badía, J. R. Cejas, F. J. Santamaría, and A. Lorenzo. 2000. Efficacy of dietary methyl esters of n-3 HUFA vs. triacylglycerols of n-3 HUFA by gilthead seabream (*Sparus aurata* L.) juveniles. *Aquaculture* 190:273–287.
- Izquierdo, M. S., A. Tandler, M. Salhi, and S. Kolkovski. 2001. Influence of dietary polar lipids' quantity and quality on ingestion and assimilation of labelled fatty acids by larval gilthead seabream. *Aquaculture Nutrition* 7:153–160.
- Izquierdo, M. S., A. Obach, L. Arantzamendi, D. Montero, L. Robaina, and G. Rosenlund. 2003. Dietary lipid sources for seabream and seabass: growth performance, tissue composition and flesh quality. *Aquaculture Nutrition* 9:397–407.
- Izquierdo, M. S., D. Montero, L. Robaina, M. J. Caballero, G. Rosenlund, and R. Ginés. 2005. Alterations in fillet fatty acid profile and flesh quality in gilthead seabream (*Sparus aurata*) fed vegetable oils for a long term period. Recovery of fatty acid profiles by fish oil feeding. *Aquaculture* 250:431–444.
- Izquierdo, M. S., L. Robaina, E. Juárez-Carrillo, V. Oliva, C. M. Hernández-Cruz, and J. M. Afonso. 2008. Regulation of growth, fatty acid composition and delta 6 desaturase expression by dietary lipids in gilthead seabream larvae (*Sparus aurata*). *Fish Physiology and Biochemistry* 34:117–127.
- Jalali Haji-abadi, S. M. A., N. Mahboobi Soofiani, A. A. Sadeghi, M. Chamani, and G. H. Riaz. 2010. Effects of supplemental dietary l-carnitine and ractopamine on the performance of juvenile rainbow trout, *Oncorhynchus mykiss*. *Aquaculture Research* 41:1582–1591.

- Jankowska, B., Z. Zakęś, T. Żmijewski, and M. Szczepkowski. 2008. Fatty acid composition of wild and cultured northern pike (*Esox lucius*). *Journal of Applied Ichthyology* 24:196–201.
- Jaya-Ram, A., M.-K. Kuah, P.-S. Lim, S. Kolkovski, and A. C. Shu-Chien. 2008. Influence of dietary HUFA levels on reproductive performance, tissue fatty acid profile and desaturase and elongase mRNAs expression in female zebrafish *Danio rerio*. *Aquaculture* 277:275–281.
- Jeong, B.-Y., S.-K. Moon, W.-G. Jeong, and T. Ohshima. 2000. Lipid classes and fatty acid compositions of wild and cultured sweet smelt *Plecoglossus altivelis* muscles and eggs in Korea. *Fisheries Science* 66:716–724.
- Jeong, D.-S., Y. Kayano, T. Oda, and H. Nakagawa. 2003. Influence of feeding regime on fatty acid composition in young red-spotted grouper *Epinephelus akaara*. *Fisheries Science* 69:569–574.
- Ji, H., A. D. Om, T. Umino, H. Nakagawa, T. Sasaki, K. Okada, M. Asano, and A. Nakagawa. 2003. Effect of dietary ascorbate fortification on lipolysis activity of juvenile black sea bream *Acanthopagrus schlegelii*. *Fisheries Science* 69:66–73.
- Ji, S.-C., G.-S. Jeong, G.-S. Im, S.-W. Lee, J.-H. Yoo, and K. Takii. 2007. Dietary medicinal herbs improve growth performance, fatty acid utilization, and stress recovery of Japanese flounder. *Fisheries Science* 73:70–76.
- Ji, S.-C., O. Takaoka, A. K. Biswas, M. Seoka, K. Ozaki, J. Kohbara, M. Ukawa, S. Shimeno, H. Hosokawa, and K. Takii. 2008. Dietary utility of enzyme-treated fish meal for juvenile Pacific bluefin tuna *Thunnus orientalis*. *Fisheries Science* 74:54–61.
- Jobling, M., A. V. Larsen, B. Andreassen, T. Sigholt, and R. L. Olsen. 2002. Influence of a dietary shift on temporal changes in fat deposition and fatty acid composition of Atlantic salmon post-smolt during the early phase of seawater rearing. *Aquaculture Research* 33:875–889.
- Jobling, M., O. Leknes, B.-S. Sæther, and E. Å. Bendiksen. 2008. Lipid and fatty acid dynamics in Atlantic cod, *Gadus morhua*, tissues: Influence of dietary lipid concentrations and feed oil sources. *Aquaculture* 281:87–94.
- Kalla, A., T. Yoshimatsu, T. Araki, D.-M. Zhang, T. Yamamoto, and S. Sakamoto. 2008. Use of *Porphyra* spheroplasts as feed additive for red sea bream. *Fisheries Science* 74:104–108.
- Kamler, E., J. Wolnicki, R. Kamiński, and J. Sikorska. 2008. Fatty acid composition, growth and morphological deformities in juvenile cyprinid, *Scardinius erythrophthalmus* fed formulated diet supplemented with natural food. *Aquaculture* 278:69–76.
- Karakatsouli, N., S. E. Papoutsoglou, G. Pizzonia, G. Tsatsos, A. Tsopekos, S. Chadio, D. Kalogiannis, C. Dalla, A. Polissidis, and Z. Papadopoulou-Daifoti. 2007. Effects of light spectrum on growth and physiological status of gilthead seabream *Sparus aurata* and rainbow trout *Oncorhynchus mykiss* reared under recirculating system conditions. *Aquacultural Engineering* 36:302–309.
- Karalazos, V., E. Å. Bendiksen, J. R. Dick, and J. G. Bell. 2007. Effects of dietary protein, and fat level and rapeseed oil on growth and tissue fatty acid composition and metabolism in Atlantic salmon (*Salmo salar* L.) reared at low water temperatures. *Aquaculture Nutrition* 13:256–265.
- Kasper, C. S., B. A. Watkins, and P. B. Brown. 2007. Evaluation of two soybean meals fed to yellow perch (*Perca flavescens*). *Aquaculture Nutrition* 13:431–438.
- Kennedy, S. R., R. Bickerdike, R. K. Berge, A. R. Porter, and D. R. Tocher. 2007a. Influence of dietary conjugated linoleic acid (CLA) and tetradecylthioacetic acid (TTA) on growth, lipid composition and key enzymes of fatty acid oxidation in liver and muscle of Atlantic cod (*Gadus morhua* L.). *Aquaculture* 264:372–382.
- Kennedy, S. R., R. Bickerdike, R. K. Berge, J. R. Dick, and D. R. Tocher. 2007b. Influence of conjugated linoleic acid (CLA) or tetradecylthioacetic acid (TTA) on growth, lipid composition, fatty acid metabolism and lipid gene expression of rainbow trout (*Oncorhynchus mykiss* L.). *Aquaculture* 272:489–501.
- Kestemont, P., E. Vandeloise, C. Mélard, P. Fontaine, and P. B. Brown. 2001. Growth and nutritional status of Eurasian perch *Perca fluviatilis* fed graded levels of dietary lipids with or without added ethoxyquin. *Aquaculture* 203:85–99.
- Khan, M. N. D., T. Yoshimatsu, A. Kalla, T. Araki, and S. Sakamoto. 2008. Supplemental effect of *Porphyra* spheroplasts on the growth and feed utilization of black sea bream. *Fisheries Science* 74:397–404.
- Khérifi, S., M. El Cafsi, W. Masmoudi, J. D. Castell, and M. S. Romdhane. 2003. Salinity and temperature effects on the lipid composition of mullet sea fry (*Mugil cephalus*, Linne, 1758). *Aquaculture International* 11:571–582.
- Khozin-Goldberg, I., Z. Cohen, M. Pimenta-Leibowitz, J. Nechev, and D. Zilberg. 2006. Feeding with arachidonic acid-rich triacylglycerols from the microalga *Parietochloris incisa* improved recovery of guppies from infection with *Tetrahymena* sp. *Aquaculture* 255:142–150.
- Kiessling, A., J. Pickova, J. G. Eales, B. Dosanjh, and D. Higgs. 2005. Age, ration level, and exercise affect the fatty acid profile of chinook salmon (*Oncorhynchus tshawytscha*) muscle differently. *Aquaculture* 243:345–356.
- Kikuchi, K., T. Furuta, N. Iwata, K. Onuki, and T. Noguchi. 2009. Effect of dietary lipid levels on the growth, feed utilization, body composition and blood characteristics of tiger puffer *Takifugu rubripes*. *Aquaculture* 298:111–117.
- Kim, K.-D. and S.-M. Lee. 2004. Requirement of dietary n-3 highly unsaturated fatty acids for juvenile flounder (*Paralichthys olivaceus*). *Aquaculture* 229:315–323.
- Kjørsvik, E., C. Olsen, P.-A. Wold, K. Hoehne-Reitan, C. L. Cahu, J. Rainuzzo, A. I. Olsen, G. Øie, and

- Y. Olsen.** 2009. Comparison of dietary phospholipids and neutral lipids on skeletal development and fatty acid composition in Atlantic cod (*Gadus morhua*). *Aquaculture* 294:246–255.
- Koizumi, K. and S. Hiratsuka.** 2009. Fatty acid compositions in muscles of wild and cultured ocellate puffer *Takifugu rubripes*. *Fisheries Science* 75:1323–1328.
- Kolakowska, A., M. Szczygielski, G. Bienkiewicz, and L. Zienkiewicz.** 2000. Some of fish species as a source of n-3 polyunsaturated fatty acids. *Acta Ichthyologica et Piscatoria* 30:59–70.
- Kolkowski, S., S. Czesny, C. Yackey, R. Moreau, F. Cihla, D. Mahan, and K. Dabrowski.** 2000. The effect of vitamins C and E in (n-3) highly unsaturated fatty acids-enriched *Artemia nauplii* on growth, survival, and stress resistance of fresh water wall-eye *Stizostedion vitreum* larvae. *Aquaculture Nutrition* 6:199–206.
- Koven, W., Y. Barr, S. Lutzky, I. Ben-Atia, R. Weiss, M. Harel, P. Behrens, and A. Tandler.** 2001. The effect of dietary arachidonic acid (20:4n-6) on growth, survival and resistance to handling stress in gilthead seabream (*Sparus aurata*) larvae. *Aquaculture* 193:107–122.
- Koven, W., R. van Anholt, S. Lutzky, I. Ben Atia, O. Nixon, B. Ron, and A. Tandler.** 2003. The effect of dietary arachidonic acid on growth, survival, and cortisol levels in different-age gilthead seabream larvae (*Sparus auratus*) exposed to handling or daily salinity change. *Aquaculture* 228:307–320.
- Kowalska, A., Z. Zakeš, B. Jankowska, and A. Siwicki.** 2010. Impact of diets with vegetable oils on the growth, histological structure of internal organs, biochemical blood parameters, and proximate composition of pikeperch *Sander lucioperca* (L.). *Aquaculture* 301:69–77.
- Kowalska, A., Z. Zakeš, B. Jankowska, and A. Siwicki.** 2011. Substituting vegetable oil for fish oil in pikeperch diets: the impact on growth, internal organ histology, blood biochemical parameters, and proximate composition. *Aquaculture Nutrition* 17: e148–e163.
- Kucska, B., L. Pál, T. Müller, M. Bódis, Á. Bartos, L. Wágner, F. Husvéth, and M. Bercsényi.** 2006. Changing of fat content and fatty acid profile of reared pike (*Esox lucius*) fed two different diets. *Aquaculture Research* 37:96–101.
- Lane, R. L. and C. C. Kohler.** 2007. Influence of organic fertilizer source on fatty acid composition of zooplankton and sunshine bass fingerlings. *North American Journal of Aquaculture* 69:413–418.
- Lee, S.-M. and K.-D. Kim.** 2005. Effect of various levels of lipid exchanged with dextrin at different protein level in diet on growth and body composition of juvenile flounder *Paralichthys olivaceus*. *Aquaculture Nutrition* 11:435–442.
- Lee, S.-M. and K.-D. Kim.** 2009. Effects of dietary carbohydrate to lipid ratios on growth and body composition of juvenile and grower rockfish, *Sebastes schlegeli*. *Aquaculture Research* 40:1830–1837.
- Lee, S.-M. and S. H. Cho.** 2009. Influences of dietary fatty acid profile on growth, body composition and blood chemistry in juvenile fat cod (*Hexagrammos otakii* Jordan et Starks). *Aquaculture Nutrition* 15:19–28.
- Lee, S.-M., S. H. Cho, and K.-D. Kim.** 2000. Effects of dietary protein and energy levels on growth and body composition of juvenile flounder *Paralichthys olivaceus*. *Journal of the World Aquaculture Society* 31:306–315.
- Lee, S.-M., J. H. Lee, and K.-D. Kim.** 2003. Effect of dietary essential fatty acids on growth, body composition and blood chemistry of juvenile starry flounder (*Platichthys stellatus*). *Aquaculture* 225:269–281.
- Lewis, H. A. and C. C. Kohler.** 2008a. Corn gluten meal partially replaces dietary fish meal without compromising growth or fatty acid composition of sunshine bass. *North American Journal of Aquaculture* 70:50–60.
- Lewis, H. A. and C. C. Kohler.** 2008b. Minimizing fish oil and fish meal with plant-based alternatives in sunshine bass diets without negatively impacting growth and muscle fatty acid profile. *Journal of the World Aquaculture Society* 39:573–585.
- Lewis-McCrea, L. M. and S. P. Lall.** 2007. Effects of moderately oxidized dietary lipid and the role of vitamin E on the development of skeletal abnormalities in juvenile Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture* 262:142–155.
- Li, Y.-Y., W.-Z. Chen, Z.-W. Sun, J.-H. Chen, and K.-G. Wu.** 2005. Effects of n-3 HUFA content in broodstock diet on spawning performance and fatty acid composition of eggs and larvae in *Plectorhynchus cinctus*. *Aquaculture* 245:263–272.
- Li, M. H., E. H. Robinson, C. S. Tucker, B. B. Manning, and L. Khoo.** 2009. Effects of dried algae *Schizochytrium* sp., a rich source of docosahexaenoic acid, on growth, fatty acid composition, and sensory quality of channel catfish *Ictalurus punctatus*. *Aquaculture* 292:232–236.
- Lim, C., M. Yildirim-Aksoy, R. Shelby, M. H. Li, and P. H. Klesius.** 2010. Growth performance, vitamin E status, and proximate and fatty acid composition of channel catfish, *Ictalurus punctatus*, fed diets containing various levels of fish oil and vitamin E. *Fish Physiology and Biochemistry* 36:855–866.
- Lin, Y.-H. and S.-Y. Shiau.** 2007. Effects of dietary blend of fish oil with corn oil on growth and non-specific immune responses of grouper, *Epinephelus malabaricus*. *Aquaculture Nutrition* 13:137–144.
- Lin, H.-Z., Y.-J. Liu, J.-G. He, W.-H. Zheng, and L. -X. Tian.** 2007. Alternative vegetable lipid sources in diets for grouper, *Epinephelus coioides* (Hamilton): effects on growth, and muscle and liver fatty acid composition. *Aquaculture Research* 38:1605–1611.
- Lin, Q., J. Lin and C. Wang.** 2009. Biochemical composition of the wild and cultured seahorses, *Hippocampus*

- kuda* Bleeker and *Hippocampus trimaculatus* Leach. *Aquaculture Research* 40:710–719.
- Ling, S., M.-K. Kuah, T. S. Tengku Muhammad, S. Kolkovski, and A. C. Shu-Chien. 2006. Effect of dietary HUFA on reproductive performance, tissue fatty acid profile and desaturase and elongase mRNAs in female swordtail *Xiphophorus helleri*. *Aquaculture* 261:204–214.
- Liu, J., M. J. Caballero, M. Izquierdo, T. El-Sayed Ali, C. M. Hernández-Cruz, A. Valencia, and H. Fernández-Palacios. 2002. Necessity of dietary lecithin and eicosapentaenoic acid for growth, survival, stress resistance and lipoprotein formation in gilthead sea bream *Sparus aurata*. *Fisheries Science* 68:1165–1172.
- López, L. M., E. Durazo, M. T. Viana, M. Drawbridge, and D. P. Bureau. 2009. Effect of dietary lipid levels on performance, body composition and fatty acid profile of juvenile white seabass, *Atractoscion nobilis*. *Aquaculture* 289:101–105.
- Lu, J., T. Takeuchi, and H. Ogawa. 2003. Flesh quality of tilapia *Oreochromis niloticus* fed solely on raw *Spirulina*. *Fisheries Science* 69:529–534.
- Ludwig, G. M., S. D. Rawles, and S. E. Lochmann. 2008. Effect of rotifer enrichment on sunshine bass *Morone chrysops* × *M. saxatilis* larvae growth and survival and fatty acid composition. *Journal of the World Aquaculture Society* 39:158–173.
- Lund, I. and S. J. Steenfeldt. 2011. The effects of dietary long-chain essential fatty acids on growth and stress tolerance in pikeperch larvae (*Sander lucioperca* L.). *Aquaculture Nutrition* 17:191–199.
- Lund, I., S. J. Steenfeldt, and B. W. Hansen. 2007. Effect of dietary arachidonic acid, eicosapentaenoic acid and docosahexaenoic acid on survival, growth and pigmentation in larvae of common sole (*Solea solea* L.). *Aquaculture* 273:532–544.
- Lund, I., S. J. Steenfeldt, G. Banta, and B. W. Hansen. 2008a. The influence of dietary concentrations of arachidonic acid and eicosapentaenoic acid at various stages of larval ontogeny on eye migration, pigmentation and prostaglandin content of common sole larvae (*Solea solea* L.). *Aquaculture* 276:143–153.
- Lund, I., S. J. Steenfeldt, K. I. Suhr, and B. W. Hansen. 2008b. A comparison of fatty acid composition and quality aspects of eggs and larvae from cultured and wild broodstock of common sole (*Solea solea* L.). *Aquaculture Nutrition* 14:544–555.
- Lund, I., S. J. Steenfeldt, and B. W. Hansen. 2010. Influence of dietary arachidonic acid combined with light intensity and tank colour on pigmentation of common sole (*Solea solea* L.) larvae. *Aquaculture* 308:159–165.
- Luo, Z., X. Y. Tan, W. M. Wang, and Q. X. Fan. 2009. Effects of long-term starvation on body weight and body composition of juvenile channel catfish, *Ictalurus punctatus*, with special emphasis on amino acid and fatty acid changes. *Journal of Applied Ichthyology* 25(2):184–189.
- Luo, G., J. Xu, Y. Teng, C. Ding, and B. Yan. 2010. Effects of dietary lipid levels on the growth, digestive enzyme, feed utilization and fatty acid composition of Japanese sea bass (*Lateolabrax japonicus* L.) reared in freshwater. *Aquaculture Research* 41:210–219.
- Luzzana, U., M. Scolari, B. Campo Dall'Orto, F. A. Vaini, N. Nargaye, and F. Valfrè. 2002. Fillet yield and chemical composition of farm-raised sunshine bass (*Morone chrysops* ♀ × *Morone saxatilis* ♂) fed high-energy diets. *Journal of Applied Ichthyology* 18:65–69.
- Luzzana, U., M. Scolari, B. Campo Dall'Orto, F. Caprino, G. Turchini, E. Orban, F. Sinesio, and F. Valfrè. 2003. Growth and product quality of European eel (*Anguilla anguilla*) as affected by dietary protein and lipid sources. *Journal of Applied Ichthyology* 19:74–78.
- Maina, J. G., R. M. Beames, D. Higgs, P. N. Mbugua, G. Iwama, and S. M. Kisia. 2003. Partial replacement of fishmeal with sunflower cake and corn oil in diets for tilapia *Oreochromis niloticus* (Linn): effect on whole body fatty acids. *Aquaculture Research* 34:601–608.
- Mairesse, G., M. Thomas, J.-N. Gardeur, and J. Brun-Bellut. 2007. Effects of dietary factors, stocking biomass and domestication on the nutritional and technological quality of the Eurasian perch *Perca fluviatilis*. *Aquaculture* 262:86–94.
- Manning, B. B., M. H. Li, E. H. Robinson, and B. C. Peterson. 2006. Enrichment of channel catfish (*Ictalurus punctatus*) filets with conjugated linoleic acid and omega-3 fatty acids by dietary manipulation. *Aquaculture* 261:337–342.
- Manning, B. B., M. H. Li, and E. H. Robinson. 2007. Feeding channel catfish, *Ictalurus punctatus*, diets amended with refined marine fish oil elevates omega-3 highly unsaturated fatty acids in filets. *Journal of the World Aquaculture Society* 38:49–58.
- Maranesi, M., M. Marchetti, D. Boichicchio, and L. Cabrini. 2005. Vitamin B6 supplementation increases the docosahexaenoic acid concentration of muscle lipids of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture Research* 36:431–438.
- Mares, J., M. Palikova, R. Kopp, S. Navratil, and J. Pikula. 2009. Changes in the nutritional parameters of muscles of the common carp (*Cyprinus carpio*) and the silver carp (*Hypophthalmichthys molitrix*) following environmental exposure to cyanobacterial water bloom. *Aquaculture Research* 40:148–156.
- Martín, M. V., C. Rodríguez, J. R. Cejas, M. J. Pérez, S. Jerez, and A. Lorenzo. 2009. Body lipid and fatty acid composition in male gilthead seabream broodstock at different stages of the reproductive cycle: effects of a diet lacking n-3 and n-6 HUFA. *Aquaculture Nutrition* 15:60–72.
- Martínez-Llorens, S., A. T. Vidal, A. V. Moñino, M. P. Torres, and M. J. Cerdá. 2007. Effects of dietary soybean oil concentration on growth, nutrient utilization and muscle fatty acid composition of

- gilthead sea bream (*Sparus aurata* L.). *Aquaculture Research* 38:76–81.
- Martino, R. C., J. E. P. Cyrino, L. Portz, and L. C. Trugo.** 2002. Performance and fatty acid composition of surubim (*Pseudoplatystoma coruscans*) fed diets with animal and plant lipids. *Aquaculture* 209:233–246.
- Martino, R. C., L. C. Trugo, J. E. P. Cyrino, and L. Portz.** 2003. Use of white fat as a replacement for squid liver oil in practical diets for surubim *Pseudoplatystoma coruscans*. *Journal of the World Aquaculture Society* 34:192–202.
- Martins, D. A., L. M. P. Valente, and S. P. Lall.** 2007. Effects of dietary lipid level on growth and lipid utilization by juvenile Atlantic halibut (*Hippoglossus hippoglossus*, L.). *Aquaculture* 263:150–158.
- Masuda, R., D. A. Ziemann, and A. C. Ostrowski.** 2001. Patchiness formation and development of schooling behavior in Pacific threadfin *Polydactylus sexfilis* reared with different dietary highly unsaturated fatty acid contents. *Journal of the World Aquaculture Society* 32:309–316.
- McKenzie, D. J., G. Piraccini, M. Piccolella, J. F. Steffensen, C. L. Bolis, and E. W. Taylor.** 2000. Effects of dietary fatty acid composition on metabolic rate and responses to hypoxia in the European eel (*Anguilla anguilla*). *Fish Physiology and Biochemistry* 22:281–296.
- Menoyo, D., C. J. Lopez-Bote, J. M. Bautista, and A. Obach.** 2002. Herring vs. anchovy oils in salmon feeding. *Aquatic Living Resources* 15:217–223.
- Menoyo, D., C. J. Lopez-Bote, J. M. Bautista, and A. Obach.** 2003. Growth, digestibility and fatty acid utilization in large Atlantic salmon (*Salmo salar*) fed varying levels of n-3 and saturated fatty acids. *Aquaculture* 225:295–307.
- Menoyo, D., C. J. Lopez-Bote, A. Diez, A. Obach, and J. M. Bautista.** 2007. Impact of n-3 fatty acid chain length and n-3/n-6 ratio in Atlantic salmon (*Salmo salar*) diets. *Aquaculture* 267:248–259.
- Mercier, L., C. Audet, J. de la Noüe, B. Parent, C. C. Parrish, and N. W. Ross.** 2004. First feeding of winter flounder (*Pseudopleuronectes americanus*) larvae: use of *Brachionus plicatilis* acclimated at low temperature as live prey. *Aquaculture* 229:361–376.
- Mishra, K. and K. Samantaray.** 2004. Interacting effects of dietary lipid level and temperature on growth, body composition and fatty acid profile of rohu, *Labeo rohita* (Hamilton). *Aquaculture Nutrition* 10:359–369.
- Mohd-Yusof, N. Y., O. Monroig, A. Mohd-Adnan, K.-L. Wan, and D. R. Tocher.** 2010. Investigation of highly unsaturated fatty acid metabolism in the Asian sea bass, *Lates calcarifer*. *Fish Physiology and Biochemistry* 36:827–843.
- Molnár, T., A. Szabó, G. Szabó, C. Szabó, and C. Hancz.** 2006. Effect of different dietary fat content and fat type on the growth and body composition of intensively reared pikeperch *Sander lucioperca* (L.). *Aquaculture Nutrition* 12:173–182.
- Monroig, Ó., J. C. Navarro, F. Amat, P. González, A. Bermejo, and F. Hontoria.** 2006. Enrichment of *Artemia* nauplii in essential fatty acids with different types of liposomes and their use in the rearing of gilthead sea bream (*Sparus aurata*) larvae. *Aquaculture* 251:491–508.
- Montero, D., L. E. Robaina, J. Socorro, J. M. Vergara, L. Tort, and M. S. Izquierdo.** 2001. Alteration of liver and muscle fatty acid composition in gilthead seabream (*Sparus aurata*) juveniles held at high stocking density and fed an essential fatty acid deficient diet. *Fish Physiology and Biochemistry* 24:63–72.
- Montero, D., L. Robaina, M. J. Caballero, R. Ginés, and M. S. Izquierdo.** 2005. Growth, feed utilization and flesh quality of European sea bass (*Dicentrarchus labrax*) fed diets containing vegetable oils: A time-course study on the effect of a re-feeding period with a 100% fish oil diet. *Aquaculture* 248:121–134.
- Montero, D., G. Lalumera, M. S. Izquierdo, M. J. Caballero, M. Saroglia, and L. Tort.** 2009. Establishment of dominance relationships in gilthead sea bream *Sparus aurata* juveniles during feeding: effects on feeding behaviour, feed utilization and fish health. *Journal of Fish Biology* 74:790–805.
- Morais, S., J. G. Bell, D. A. Robertson, W. J. Roy, and P. C. Morris.** 2001. Protein/lipid ratios in extruded diets for Atlantic cod (*Gadus morhua* L.): effects on growth, feed utilisation, muscle composition and liver histology. *Aquaculture* 203:101–119.
- Morais, S., L. Narciso, E. Dores, and P. Pousão-Ferreira.** 2004. Lipid enrichment for Senegalese sole (*Solea senegalensis*) larvae: effect on larval growth, survival and fatty acid profile. *Aquaculture International* 12:281–298.
- Mørkøre, T.** 2006. Relevance of dietary oil source for contraction and quality of pre-rigor filleted Atlantic cod, *Gadus morhua*. *Aquaculture* 251:56–65.
- Mørkøre, T., C. Netteberg, L. Johnsson, and J. Pickova.** 2007. Impact of dietary oil source on product quality of farmed Atlantic cod, *Gadus morhua*. *Aquaculture* 267:236–247.
- Morris, P. C., P. Gallimore, J. Handley, G. Hide, P. Haughton, and A. Black.** 2005. Full-fat soya for rainbow trout (*Oncorhynchus mykiss*) in freshwater: Effects on performance, composition and flesh fatty acid profile in absence of hind-gut enteritis. *Aquaculture* 248:147–161.
- Mourete, G., J. E. Good, and J. G. Bell.** 2005. Partial substitution of fish oil with rapeseed, linseed and olive oils in diets for European sea bass (*Dicentrarchus labrax* L.): effects on flesh fatty acid composition, plasma prostaglandins E₂ and F_{2α}, immune function and effectiveness of a fish oil finishing diet. *Aquaculture Nutrition* 11:25–40.
- Mráz, J. and J. Pickova.** 2009. Differences between lipid content and composition of different parts of fillets from crossbred farmed carp (*Cyprinus carpio*). *Fish Physiology and Biochemistry* 35:615–623.

- Mráz, J., C. Schlechtriem, L. A. Olohan, Y. Fang, A. R. Cossins, V. Zlabek, T. Samuelsen, and J. Pickova. 2010. Sesamin as a potential modulator of fatty acid composition in common carp (*Cyprinus carpio*). *Aquaculture Research* 41:e851–e861.
- Nakagawa, H., M. Furuhashi, T. Umino, A. Takago, and S. Sakamoto. 2004. Utilization of α -starch in ayu, *Plecoglossus altivelis*, relating to growth and body composition. *Journal of Applied Ichthyology* 20:389–394.
- Nandi, S., P. Routray, S. D. Gupta, S. C. Rath, S. Dasgupta, P. K. Meher, and P. K. Mukhopadhyay. 2007. Reproductive performance of carp, *Catla catla* (Ham.), reared on a formulated diet with PUFA supplementation. *Journal of Applied Ichthyology* 23:684–691.
- Nanton, D. A., S. P. Lall, and M. A. McNiven. 2001. Effects of dietary lipid level on liver and muscle lipid deposition in juvenile haddock, *Melanogrammus aeglefinus* L. *Aquaculture Research* 32:225–234.
- Nanton, D. A., A. Vegusdal, A. M. B. Rørå, B. Ruyter, G. Bæverfjord, and B. E. Torstensen. 2007. Muscle lipid storage pattern, composition, and adipocyte distribution in different parts of Atlantic salmon (*Salmo salar*) fed fish oil and vegetable oil. *Aquaculture* 265:230–243.
- Naz, M. 2008. Ontogeny of fertilized eggs and yolk sac larvae of sea bass (*Dicentrarchus labrax*). *The Israeli Journal of Aquaculture* 60:113–120.
- Naz, M. 2009. Ontogeny of biochemical phases of fertilized eggs and yolk sac larvae of gilthead seabream (*Sparus aurata* L.). *Turkish Journal of Fisheries and Aquatic Sciences* 9:77–83.
- Ng, W. K., M. C. Tee, and P. L. Boey. 2000. Evaluation of crude palm oil and refined palm olein as dietary lipids in pelleted feeds for a tropical bagrid catfish *Mystus nemurus* (Cuvier & Valenciennes). *Aquaculture Research* 31:337–347.
- Ng, W.-K., P.-K. Lim, and H. Sidek. 2001. The influence of a dietary lipid source on growth, muscle fatty acid composition and erythrocyte osmotic fragility of hybrid tilapia. *Fish Physiology and Biochemistry* 25:301–310.
- Ng, W.-K., P.-K. Lim, and P.-L. Boey. 2003. Dietary lipid and palm oil source affects growth, fatty acid composition and muscle α -tocopherol concentration of African catfish, *Clarias gariepinus*. *Aquaculture* 215:229–243.
- Ng, W.-K., T. Sigholt, and J. G. Bell. 2004. The influence of environmental temperature on the apparent nutrient and fatty acid digestibility in Atlantic salmon (*Salmo salar* L.) fed finishing diets containing different blends of fish oil, rapeseed oil and palm oil. *Aquaculture Research* 35:1228–1237.
- Ng, W.-K., C.-B. Koh, and Z. B. Din. 2006. Palm oil-laden spent bleaching clay as a substitute for marine fish oil in the diets of Nile tilapia, *Oreochromis niloticus*. *Aquaculture Nutrition* 12:459–468.
- Nguyen, V. T., S. Satoh, Y. Haga, H. Fushimi, and T. Kotani. 2008. Effect of zinc and manganese supplementation in Artemia on growth and vertebral deformity in red sea bream (*Pagrus major*) larvae. *Aquaculture* 285:184–192.
- Nielsen, N. S., J. R. Göttische, J. Holm, X. Xu, H. Mu, and C. Jacobsen. 2005. Effect of structured lipids based on fish oil on the growth and fatty acid composition in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 250:411–423.
- Noffs, M. D., R. C. Martino, L. C. Trugo, E. C. Urbinati, J. B. K. Fernandes, and L. S. Takahashi. 2009. Dietary fish oil replacement with lard and soybean oil affects triacylglycerol and phospholipid muscle and liver docosahexaenoic acid content but not in the brain and eyes of surubim juveniles *Pseudoplatystoma* sp. *Fish Physiology and Biochemistry* 35:399–412.
- Nogueira, N., N. Cordeiro, P. Canada, P. Cruz e Silva, and R. O. A. Ozório. 2010. Separate and combined effects of cyclic fasting and l-carnitine supplementation in red porgy (*Pagrus pagrus*, L. 1758). *Aquaculture Research* 41:e795–e806.
- Nordgarden, U., B. E. Torstensen, L. Frøyland, T. Hansen, and G. I. Hemre. 2003. Seasonally changing metabolism in Atlantic salmon (*Salmo salar* L.) II – β -oxidation capacity and fatty acid composition in muscle tissues and plasma lipoproteins. *Aquaculture Nutrition* 9:295–303.
- O'Neal, C. C. and C. C. Kohler. 2008. Effect of replacing menhaden oil with catfish oil on the fatty acid composition of juvenile channel catfish, *Ictalurus punctatus*. *Journal of the World Aquaculture Society* 39:62–71.
- Ogata, H. Y., A. C. Emata, E. S. Garibay, and H. Furuita. 2004. Fatty acid composition of five candidate aquaculture species in Central Philippines. *Aquaculture* 236:361–375.
- Ogunji, J. O. and M. Wirth. 2002. Influence of dietary protein deficiency on amino acid and fatty acid composition in tilapia, *Oreochromis niloticus*, fingerlings. *The Israeli Journal of Aquaculture* 54:64–72.
- Olsen, R. E., R. J. Henderson, J. Sountama, G. I. Hemre, E. Ringø, W. Melle, and D. R. Tocher. 2004. Atlantic salmon, *Salmo salar*, utilizes wax ester-rich oil from *Calanus finmarchicus* effectively. *Aquaculture* 240:433–449.
- Olsson, G. B., R. L. Olsen, M. Carlehög, and R. Ofstad. 2003. Seasonal variations in chemical and sensory characteristics of farmed and wild Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture* 217:191–205.
- Om, A. D., T. Umino, H. Nakagawa, T. Sasaki, K. Okada, M. Asano, and A. Nakagawa. 2001. The effects of dietary EPA and DHA fortification on lipolysis activity and physiological function in juvenile black sea bream *Acanthopagrus schlegelii* (Bleeker). *Aquaculture Research* 32:255–262.
- Ortega, A. and G. Mourente. 2010. Comparison of the lipid profiles from wild caught eggs and unfed

- larvae of two scombroid fish: northern bluefin tuna (*Thunnus thynnus* L., 1758) and Atlantic bonito (*Sarda sarda* Bloch, 1793). *Fish Physiology and Biochemistry* 36:461–471.
- Ostaszewska, T., A. Boruta, and M. Olejniczak.** 2005. The effect of dietary lipid level and composition on growth, survival, and development of the digestive system of larval sneep, *Chondrostoma nasus* (L.). *Acta Ichthyologica et Piscatoria* 35:79–86.
- Østbye, T. K., M. A. Kjaer, A. M. B. Rørå, B. Torstensen, and B. Ruyter.** 2011. High n-3 HUFA levels in the diet of Atlantic salmon affect muscle and mitochondrial membrane lipids and their susceptibility to oxidative stress. *Aquaculture Nutrition* 17:177–190.
- Ozaki, Y., H. Koga, T. Takahashi, S. Adachi, and K. Yamauchi.** 2008. Lipid content and fatty acid composition of muscle, liver, ovary and eggs of captive-reared and wild silver Japanese eel *Anguilla japonica* during artificial maturation. *Fisheries Science* 74:362–371.
- Palmegiano, G. B., E. Agradi, G. Forneris, F. Gai, L. Gasco, E. Rigamonti, B. Sicuro, and I. Zoccarato.** 2005. Spirulina as a nutrient source in diets for growing sturgeon (*Acipenser baeri*). *Aquaculture Research* 36:188–195.
- Palmegiano, G. B., F. Daprà, G. Forneris, F. Gai, L. Gasco, K. Guo, P. G. Peiretti, B. Sicuro, and I. Zoccarato.** 2006. Rice protein concentrate meal as a potential ingredient in practical diets for rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* 258:357–367.
- Palmegiano, G. B., F. Gai, F. Daprà, L. Gasco, M. Pazzaglia, and P. G. Peiretti.** 2008. Effects of Spirulina and plant oil on the growth and lipid traits of white sturgeon (*Acipenser transmontanus*) fingerlings. *Aquaculture Research* 39:587–595.
- Palmeri, G., G. M. Turchini, P. J. Marriott, P. Morrison, and S. S. De Silva.** 2009a. Biometric, nutritional and sensory characteristic modifications in farmed Murray cod (*Maccullochella peelii peelii*) during the purging process. *Aquaculture* 287:354–360.
- Palmeri, G., G. M. Turchini, and S. S. De Silva.** 2009b. Short-term food deprivation does not improve the efficacy of a fish oil finishing strategy in Murray cod. *Aquaculture Nutrition* 15:657–666.
- Papoutsoglou, S. E., N. Karakatsouli, and P. Koustas.** 2005. Effects of dietary l-tryptophan and lighting conditions on growth performance of European sea bass (*Dicentrarchus labrax*) juveniles reared in a recirculating water system. *Journal of Applied Ichthyology* 21:520–524.
- Papoutsoglou, S. E., N. Karakatsouli, E. Louizos, S. Chadio, D. Kalogiannis, C. Dalla, A. Polissidis, and Z. Papadopoulou-Daifoti.** 2007. Effect of Mozart's music (Romanze-Andante of "Eine Kleine Nacht Musik," sol major, K525) stimulus on common carp (*Cyprinus carpio* L.) physiology under different light conditions. *Aquacultural Engineering* 36:61–72.
- Papoutsoglou, S. E., N. Karakatsouli, A. Batzina, E. S. Papoutsoglou, and A. Tsopelakos.** 2008. Effect of music stimulus on gilthead seabream *Sparus aurata* physiology under different light intensity in a recirculating water system. *Journal of Fish Biology* 73:980–1004.
- Papoutsoglou, S. E., N. Karakatsouli, E. S. Papoutsoglou, and G. Vasilikos.** 2010. Common carp (*Cyprinus carpio*) response to two pieces of music ("Eine Kleine Nachtmusik" and "Romanza") combined with light intensity, using recirculating water system. *Fish Physiology and Biochemistry* 36:539–554.
- Park, H. G., V. Puvanendran, A. Kellett, C. C. Parrish, and J. A. Brown.** 2006. Effect of enriched rotifers on growth, survival, and composition of larval Atlantic cod (*Gadus morhua*). *ICES Journal of Marine Science* 63:285–295.
- Periago, M. J., M. D. Ayala, O. López-Albors, I. Abdel, C. Martínez, A. García-Alcázar, G. Ros, and F. Gil.** 2005. Muscle cellularity and flesh quality of wild and farmed sea bass, *Dicentrarchus labrax* L. *Aquaculture* 249:175–188.
- Person-Le Ruyet, J., A. Skalli, B. Dulau, N. Le Bayon, H. Le Delliou, and J. H. Robin.** 2004. Does dietary n-3 highly unsaturated fatty acids level influence the European sea bass (*Dicentrarchus labrax*) capacity to adapt to a high temperature? *Aquaculture* 242:571–588.
- Pettersson, A., J. Pickova, and E. Brännäs.** 2009a. Effects of crude rapeseed oil on lipid composition in Arctic charr *Salvelinus alpinus*. *Journal of Fish Biology* 75:1446–1458.
- Pettersson, A., L. Johnsson, E. Brännäs, and J. Pickova.** 2009b. Effects of rapeseed oil replacement in fish feed on lipid composition and self-selection by rainbow trout (*Oncorhynchus mykiss*). *Aquaculture Nutrition* 15:577–586.
- Pettersson, A., J. Pickova, and E. Brännäs.** 2010. Swimming performance at different temperatures and fatty acid composition of Arctic charr (*Salvelinus alpinus*) fed palm and rapeseed oils. *Aquaculture* 300:176–181.
- Piccolo, G., S. Marono, F. Bovera, R. Tudisco, G. Caricato, and A. Nizza.** 2008. Effect of stocking density and protein/fat ratio of the diet on the growth of Dover sole (*Solea solea*). *Aquaculture Research* 39:1697–1704.
- Piedecausa, M. A., M. J. Mazón, B. García García, and M. D. Hernández.** 2007. Effects of total replacement of fish oil by vegetable oils in the diets of sharpsnout seabream (*Diplodus puntazzo*). *Aquaculture* 263:211–219.
- Plante, S., F. Pernet, R. Haché, R. Ritchie, B. Ji, and D. McIntosh.** 2007. Ontogenetic variations in lipid class and fatty acid composition of had-dock larvae *Melanogrammus aeglefinus* in relation to changes in diet and microbial environment. *Aquaculture* 263:107–121.
- Poli, B. M., G. Parisi, G. Zampacavallo, F. Iurzan, M. Mecatti, P. Lupi, and A. Bonelli.** 2003. Preliminary results on quality and quality changes in reared

- meagre (*Argyrosomus regius*): body and fillet traits and freshness changes in refrigerated commercial-size fish. *Aquaculture International* 11:301–311.
- Pousão-Ferreira, P., P. Santos, A. P. Carvalho, S. Morais, and L. Narciso.** 2003. Effect of an experimental microparticulate diet on the growth, survival and fatty acid profile of gilthead seabream (*Sparus aurata* L.) larvae. *Aquaculture International* 11:491–504.
- Powell, M. S., R. W. Hardy, T. A. Flagg, and P. A. Kline.** 2010. Proximate composition and fatty acid differences in hatchery-reared and wild Snake River sockeye salmon overwintering in nursery lakes. *North American Journal of Fisheries Management* 30:530–537.
- Pratoomyot, J., E. Å. Bendiksen, J. G. Bell, and D. R. Tocher.** 2008. Comparison of effects of vegetable oils blended with southern hemisphere fish oil and decontaminated northern hemisphere fish oil on growth performance, composition and gene expression in Atlantic salmon (*Salmo salar* L.). *Aquaculture* 280:170–178.
- Pratoomyot, J., E. Å. Bendiksen, J. G. Bell, and D. R. Tocher.** 2010. Effects of increasing replacement of dietary fishmeal with plant protein sources on growth performance and body lipid composition of Atlantic salmon (*Salmo salar* L.). *Aquaculture* 305:124–132.
- Rajkumar, M. and K. P. Kumaraguru vasagam.** 2006. Suitability of the copepod, *Acartia clausi* as a live feed for Seabass larvae (*Lates calcarifer* Bloch): Compared to traditional live-food organisms with special emphasis on the nutritional value. *Aquaculture* 261:649–658.
- Ramos, A., N. M. Bandarra, P. Rema, P. Vaz-Pires, M. L. Nunes, A. M. Andrade, A. R. Cordeiro, and L. M. P. Valente.** 2008. Time course deposition of conjugated linoleic acid in market size rainbow trout (*Oncorhynchus mykiss*) muscle. *Aquaculture* 274:366–374.
- Regan, M. D., L. J. Kuchel, S. S. Y. Huang, D. A. Higgs, J. Wang, P. M. Schulte, and C. J. Brauner.** 2010. The effect of dietary fish oil and poultry fat replacement with canola oil on swimming performance and metabolic response to hypoxia in stream type spring Chinook salmon parr. *Aquaculture* 308:183–189.
- Regost, C., J. Arzel, J. Robin, G. Rosenlund, and S. J. Kaushik.** 2003. Total replacement of fish oil by soybean or linseed oil with a return to fish oil in turbot (*Psetta maxima*): 1. Growth performance, flesh fatty acid profile, and lipid metabolism. *Aquaculture* 217:465–482.
- Rennert, B., M. Wirth, S. Günther, and C. Schulz.** 2005. Effect of feeding under-year zander (*Sander lucioperca*) on size, body mass and body composition before and after wintering. *Journal of Applied Ichthyology* 21:429–432.
- Rennie, S., F. A. Huntingford, A.-L. Loeland, and M. Rimbach.** 2005. Long term partial replacement of dietary fish oil with rapeseed oil; effects on egg quality of Atlantic salmon *Salmo salar*. *Aquaculture* 248:135–146.
- Reyes, O. S., M. N. Duray, C. B. Santiago, and M. Ricci.** 2011. Growth and survival of grouper *Epinephelus coioides* (Hamilton) larvae fed free-living nematode *Panagrellus redivivus* at first feeding. *Aquaculture International* 19:155–164.
- Rezek, T. C., W. O. Watanabe, M. Harel, and P. J. Seaton.** 2010. Effects of dietary docosahexaenoic acid (22:6n-3) and arachidonic acid (20:4n-6) on the growth, survival, stress resistance and fatty acid composition in black sea bass *Centropristis striata* (Linnaeus 1758) larvae. *Aquaculture Research* 41:1302–1314.
- Rinhard, J., S. Czesny, and K. Dabrowski.** 2007. Influence of lipid class and fatty acid deficiency on survival, growth, and fatty acid composition in rainbow trout juveniles. *Aquaculture* 264:363–371.
- Robin, J. H. and A. Peron.** 2004. Consumption vs. deposition of essential fatty acids in gilthead sea bream (*Sparus aurata*) larvae fed semi-purified diets. *Aquaculture* 238:283–294.
- Robin, J. H. and A. Skalli.** 2007. Incorporation of dietary fatty acid in European sea bass (*Dicentrarchus labrax*) - A methodological approach evidencing losses of highly unsaturated fatty acids. *Aquaculture* 263:227–237.
- Robin, J. H. and B. Vincent.** 2003. Microparticulate diets as first food for gilthead sea bream larva (*Sparus aurata*): study of fatty acid incorporation. *Aquaculture* 225:463–474.
- Roncarati, A., P. Melotti, A. Dees, O. Mordenti, and L. Angellotti.** 2006. Welfare status of cultured seabass (*Dicentrarchus labrax* L.) and seabream (*Sparus aurata* L.) assessed by blood parameters and tissue characteristics. *Journal of Applied Ichthyology* 22:225–234.
- Rondán, M., M. D. Hernández, M. Á. Egea, B. García, F. M. Rueda, and F. J. Martínez.** 2004a. Effect of feeding rate on fatty acid composition of sharpsnout seabream (*Diplodus puntazzo*). *Aquaculture Nutrition* 10:301–307.
- Rondán, M., M. D. Hernández, M. A. Egea, B. García, M. Jover, F. M. Rueda, and F. J. Martínez.** 2004b. Effects of fishmeal replacement with soybean meal as protein source, and protein replacement with carbohydrates as an alternative energy source on sharpsnout sea bream, *Diplodus puntazzo*, fatty acid profile. *Aquaculture Research* 35:1220–1227.
- Roo, F. J., C. M. Hernández-Cruz, J. A. Socorro, H. Fernández-Palacios, D. Montero, and M. S. Izquierdo.** 2009a. Effect of DHA content in rotifers on the occurrence of skeletal deformities in red porgy *Pagrus pagrus* (Linnaeus, 1758). *Aquaculture* 287:84–93.
- Roo, F. J., C. M. Hernández-Cruz, J. A. Socorro, H. Fernández-Palacios, and M. S. Izquierdo.** 2010. Advances in rearing techniques of *Pagrus pagrus*,

- (Linnaeus, 1758): comparison between intensive and semi-intensive larval rearing systems. *Aquaculture Research* 41:433–449.
- Roo, J., C. M. Hernández-Cruz, C. Borrero, D. Schuchardt, and H. Fernández-Palacios. 2010b. Effect of larval density and feeding sequence on meagre (*Argyrosomus regius*; Asso, 1801) larval rearing. *Aquaculture* 302:82–88.
- Rørå, A. M. B., B. Ruyter, J. Skorve, R. K. Berge, and K.-E. Slinning. 2005. Influence of high content of dietary soybean oil on quality of large fresh, smoked and frozen Atlantic salmon (*Salmo salar*). *Aquaculture International* 13:217–231.
- Rosenlund, G., A. Obach, M. G. Sandberg, H. Standal, and K. Tveit. 2001. Effect of alternative lipid sources on long-term growth performance and quality of Atlantic salmon (*Salmo salar* L.). *Aquaculture Research* 32:323–328.
- Rueda-Jasso, R., L. E. C. Conceição, J. Dias, W. De Coen, E. Gomes, J. F. Rees, F. Soares, M. T. Dinis, and P. Sorgeloos. 2004. Effect of dietary non-protein energy levels on condition and oxidative status of Senegalese sole (*Solea senegalensis*) juveniles. *Aquaculture* 231:417–433.
- Ruyter, B., C. Røsjø, O. Einen, and M. S. Thomassen. 2000a. Essential fatty acids in Atlantic salmon: effects of increasing dietary doses of n-6 and n-3 fatty acids on growth, survival and fatty acid composition of liver, blood and carcass. *Aquaculture Nutrition* 6:119–127.
- Ruyter, B., C. Røsjø, O. Einen, and M. S. Thomassen. 2000b. Essential fatty acids in Atlantic salmon: time course of changes in fatty acid composition of liver, blood and carcass induced by a diet deficient in n-3 and n-6 fatty acids. *Aquaculture Nutrition* 6:109–117.
- Saavedra, M., P. Pousão-Ferreira, M. Yúfera, M. T. Dinis, and L. E. C. Conceição. 2009. A balanced amino acid diet improves *Diplodus sargus* larval quality and reduces nitrogen excretion. *Aquaculture Nutrition* 15:517–524.
- Saglik Aslan, S., K. C. Guven, T. Gezgin, M. Alpaslan, and A. Tekinay. 2007. Comparison of fatty acid contents of wild and cultured rainbow trout *Oncorhynchus mykiss* in Turkey. *Fisheries Science* 73:1195–1198.
- Sagstad, A., M. Sanden, Å. Krogdahl, A. M. Bakke-McKellep, M. Frøystad, and G.-I. Hemre. 2008. Organs development, gene expression and health of Atlantic salmon (*Salmo salar* L.) fed genetically modified soybeans compared to the near-isogenic non-modified parental line. *Aquaculture Nutrition* 14:556–572.
- Samuelsen, T., M. Isaksen, and E. McLean. 2001. Influence of dietary recombinant microbial lipase on performance and quality characteristics of rainbow trout, *Oncorhynchus mykiss*. *Aquaculture* 194:161–171.
- Satoh, N. and T. Takeuchi. 2009. Estimation of the period sensitive for the development of abnormal morphology in brown sole *Pseudopleuronectes herzensteini* fed live food enriched with docosahexaenoic acid. *Fisheries Science* 75:985–991.
- Satoh, N., Y. Takaya, and T. Takeuchi. 2009a. Docosahexaenoic acid requirement for the prevention of abnormal morphology in brown sole *Pseudopleuronectes herzensteini* during D-E larval stages. *Fisheries Science* 75:1259–1266.
- Satoh, N., Y. Takaya, and T. Takeuchi. 2009b. The effect of docosahexaenoic and eicosapentaenoic acids in live food on the development of abnormal morphology in hatchery-reared brown sole *Pseudopleuronectes herzensteini*. *Fisheries Science* 75:1001–1006.
- Scaife, J. R., G. E. Onibi, I. Murray, T. C. Fletcher, and D. F. Houlihan. 2000. Influence of α -tocopherol acetate on the short- and long-term storage properties of fillets from Atlantic salmon *Salmo salar* fed a high lipid diet. *Aquaculture Nutrition* 6:65–71.
- Schlechtriem, C., J. E. Bron, and D. R. Tocher. 2007. Inter-individual variation in total fatty acid compositions of flesh of Atlantic salmon smolts-fed diets containing fish oil or vegetable oil. *Aquaculture Research* 38:1045–1055.
- Schlechtriem, C., J. E. Bron, and D. R. Tocher. 2009. Determination of n-3 HUFA content in Atlantic salmon flesh based on the lipid content, morphometric measurements and blood fatty acid composition: A modeling approach. *Journal of Applied Ichthyology* 25:120–123.
- Schulz, C., U. Knaus, M. Wirth, and B. Rennert. 2005. Effects of varying dietary fatty acid profile on growth performance, fatty acid, body and tissue composition of juvenile pike perch (*Sander lucioperca*). *Aquaculture Nutrition* 11:403–413.
- Seaborn, G. T., M. L. Jahncke, and T. I. J. Smith. 2000. Differentiation between cultured hybrid striped bass and wild striped bass and hybrid bass using fatty acid profiles. *North American Journal of Fisheries Management* 20:618–626.
- Gener, E. and M. Yildiz. 2003. Effect of the different oil on growth performance and body composition of rainbow trout (*Oncorhynchus mykiss* W., 1792) juveniles. *Turkish Journal of Fisheries and Aquatic Sciences* 3:111–116.
- Seno-O, A., F. Takakuwa, T. Hashiguchi, K. Morioka, T. Masumoto, and H. Fukada. 2008. Replacement of dietary fish oil with olive oil in young yellowtail *Seriola quinqueradiata*: effects on growth, muscular fatty acid composition and prevention of dark muscle discoloration during refrigerated storage. *Fisheries Science* 74:1297–1306.
- Seoka, M., M. Kurata, and H. Kumai. 2007. Effect of docosahexaenoic acid enrichment in Artemia on growth of Pacific bluefin tuna *Thunnus orientalis* larvae. *Aquaculture* 270:193–199.
- Seoka, M., M. Kurata, R. Tamagawa, A. K. Biswas, B. K. Biswas, A. S. K. Yong, Y.-S. Kim, S.-C. Ji, K. Takii, and H. Kumai. 2008. Dietary supplementation of salmon roe phospholipid enhances the growth and survival of Pacific bluefin tuna *Thunnus orientalis* larvae and juveniles. *Aquaculture* 275:225–234.

- Seychelles, L. H., C. Audet, R. Tremblay, K. Lemarchand, and F. Pernet. 2011. Bacterial colonization of winter flounder *Pseudopleuronectes americanus* fed live feed enriched with three different commercial diets. *Aquaculture Nutrition* 17:e196–e206.
- Shapawi, R., S. Mustafa, and W.-K. Ng. 2008. Effects of dietary fish oil replacement with vegetable oils on growth and tissue fatty acid composition of humpback grouper, *Cromileptes altivelis* (Valenciennes). *Aquaculture Research* 39:315–323.
- Sharma, P., V. Kumar, A. K. Sinha, J. Ranjan, H. M. P. Kithsiri, and G. Venkateshwarlu. 2010. Comparative fatty acid profiles of wild and farmed tropical freshwater fish rohu (*Labeo rohita*). *Fish Physiology and Biochemistry* 36:411–417.
- Shields, R. J., S. Irwin, P. L. Smith, and L. A. McEvoy. 2003. Effects of diet transition regimen on survival, growth and lipid composition of intensively reared Atlantic cod, *Gadus morhua*, larvae. *Aquaculture International* 11:119–130.
- Sicuro, B., S. Barbera, F. Daprà, F. Gai, L. Gasco, G. Paglialonga, G. B. Palmegiano, and S. Vilella. 2010. The olive oil by-product in 'rainbow trout *Oncorhynchus mykiss* (Walbaum)' farming: productive results and quality of the product. *Aquaculture Research* 41:e475–e486.
- Silva, S. S. D., R. M. Gunasekera, and R. O. Collins. 2002. Some morphometric and biochemical features of ready-to-migrate silver and pre-migratory yellow stages of the shortfin eel of south-eastern Australian waters. *Journal of Fish Biology* 61:915–928.
- Skalli, A. and J. H. Robin. 2004. Requirement of n-3 long chain polyunsaturated fatty acids for European sea bass (*Dicentrarchus labrax*) juveniles: growth and fatty acid composition. *Aquaculture* 240:399–415.
- Skalli, A., J. H. Robin, N. Le Bayon, H. Le Delliou, and J. Person-Le Ruyet. 2006. Impact of essential fatty acid deficiency and temperature on tissues' fatty acid composition of European sea bass (*Dicentrarchus labrax*). *Aquaculture* 255:223–232.
- Steffens, W. and M. Wirth. 2007. Influence of nutrition on the lipid quality of pond fish: common carp (*Cyprinus carpio*) and tench (*Tinca tinca*). *Aquaculture International* 15:313–319.
- St-Hilaire, S., C. Sheppard, J. K. Tomberlin, S. Irving, L. Newton, M. A. McGuire, E. E. Mosley, R. W. Hardy, and W. Sealey. 2007. Fly prepupae as a feedstuff for rainbow trout, *Oncorhynchus mykiss*. *Journal of the World Aquaculture Society* 38:59–67.
- Stubhaug, I., Ø. Lie, and B. E. Torstensen. 2007. Fatty acid productive value and β -oxidation capacity in Atlantic salmon (*Salmo salar* L.) fed on different lipid sources along the whole growth period. *Aquaculture Nutrition* 13:145–155.
- Suárez, M. D., M. R. Cervera, E. Abellán, A. E. Morales, M. G. Gallego, and G. Cardenete. 2010. Influence of starvation on flesh quality of farmed dentex, *Dentex dentex*. *Journal of the World Aquaculture Society* 41:490–505.
- Subhadra, B., R. Lochmann, S. Rawles, and R. Chen. 2006a. Effect of dietary lipid source on the growth, tissue composition and hematological parameters of largemouth bass (*Micropterus salmoides*). *Aquaculture* 255:210–222.
- Subhadra, B., R. Lochmann, S. Rawles, and R. Chen. 2006b. Effect of fish-meal replacement with poultry by-product meal on the growth, tissue composition and hematological parameters of largemouth bass (*Micropterus salmoides*) fed diets containing different lipids. *Aquaculture* 260:221–231.
- Suja, B., H. Phillips, R. Lochmann, and R. Chen. 2009. Effect of temperature on growth, feed utilization, and immune status of channel catfish in a recirculating system. *North American Journal of Aquaculture* 71:64–72.
- Suontama, J., Ø. Karlsen, M. Moren, G. I. Hemre, W. Melle, E. Langmyhr, H. Mundheim, E. Ringø, and R. E. Olsen. 2007. Growth, feed conversion and chemical composition of Atlantic salmon (*Salmo salar* L.) and Atlantic halibut (*Hippoglossus hippoglossus* L.) fed diets supplemented with krill or amphipods. *Aquaculture Nutrition* 13:241–255.
- Sutton, J., S. Balfry, D. Higgs, C.-H. Huang, and B. Skura. 2006. Impact of iron-catalyzed dietary lipid peroxidation on growth performance, general health and flesh proximate and fatty acid composition of Atlantic salmon (*Salmo salar* L.) reared in seawater. *Aquaculture* 257:534–557.
- Szabó, A., M. Mézes, C. Hancz, T. Molnár, D. Varga, R. Romvári, and H. Félbel. 2011. Incorporation dynamics of dietary vegetable oil fatty acids into the triacylglycerols and phospholipids of tilapia (*Oreochromis niloticus*) tissues (fillet, liver, visceral fat and gonads). *Aquaculture Nutrition* 17:e132–e147.
- Takeuchi, T., J. U. N. Lu, G. Yoshizaki, and S. Satoh. 2002. Effect on the growth and body composition of juvenile tilapia *Oreochromis niloticus* fed raw *Spirulina*. *Fisheries Science* 68:34–40.
- Tidwell, J. H., S. Coyle, and L. A. Bright. 2007. Effects of different types of dietary lipids on growth and fatty acid composition of largemouth bass. *North American Journal of Aquaculture* 69:257–264.
- Tocher, D. R., J. G. Bell, J. R. Dick, R. J. Henderson, F. McGhee, D. Michell, and P. C. Morris. 2000. Polyunsaturated fatty acid metabolism in Atlantic salmon (*Salmo salar*) undergoing parr-smolt transformation and the effects of dietary linseed and rapeseed oils. *Fish Physiology and Biochemistry* 23:59–73.
- Tomita, Y. and Y. Ando. 2009. Reinvestigation of positional distribution of tetracosahexaenoic acid in triacyl-sn-glycerols of flathead flounder flesh. *Fisheries Science* 75:445–451.
- Tonial, I. B., F. B. Stevanato, M. Matsushita, N. E. De Souza, W. M. Furuya, and J. V. Visentainer. 2009. Optimization of flaxseed oil feeding time length in adult Nile tilapia (*Oreochromis niloticus*) as a function of muscle omega-3 fatty acids composition. *Aquaculture Nutrition* 15:564–568.

- Torstensen, B. E., Ø. Lie, and K. Hamre. 2001. A factorial experimental design for investigation of effects of dietary lipid content and pro- and antioxidants on lipid composition in Atlantic salmon (*Salmo salar*) tissues and lipoproteins. *Aquaculture Nutrition* 7:265–276.
- Torstensen, B. E., L. Frøyland, and Ø. Lie. 2004. Replacing dietary fish oil with increasing levels of rapeseed oil and olive oil – effects on Atlantic salmon (*Salmo salar* L.) tissue and lipoprotein lipid composition and lipogenic enzyme activities. *Aquaculture Nutrition* 10:175–192.
- Trattner, S., J. Pickova, K. H. Park, J. Rinchar, and K. Dabrowski. 2007. Effects of [alpha]-lipoic and ascorbic acid on the muscle and brain fatty acids and antioxidant profile of the South American pacu *Piaractus mesopotamicus*. *Aquaculture* 273:158–164.
- Trenzado, C. E., A. E. Morales, and M. de la Higuera. 2008. Physiological changes in rainbow trout held under crowded conditions and fed diets with different levels of vitamins E and C and highly unsaturated fatty acids (HUFA). *Aquaculture* 277:293–302.
- Trushenski, J. T. 2009. Saturated lipid sources in feeds for sunshine bass: alterations in production performance and tissue fatty acid composition. *North American Journal of Aquaculture* 71:363–373.
- Trushenski, J. T. and J. Boesenberg. 2009. Influence of dietary fish oil concentration and finishing duration on beneficial fatty acid profile restoration in sunshine bass *Morone chrysops* ♀ x *M. saxatilis* ♂. *Aquaculture* 296:277–283.
- Trushenski, J. T. and C. C. Kohler. 2008. Influence of stress, exertion, and dietary natural source vitamin E on prostaglandin synthesis, hematology, and tissue fatty acid composition of sunshine bass. *North American Journal of Aquaculture* 70:251–265.
- Trushenski, J. T., J. Boesenberg, and C. C. Kohler. 2009. Influence of grow-out feed fatty acid composition on finishing success in Nile tilapia. *North American Journal of Aquaculture* 71:242–251.
- Trushenski, J., M. Schwarz, H. Lewis, J. Laporte, B. Delbos, R. Takeuchi, and L. A. Sampaio. 2011. Effect of replacing dietary fish oil with soybean oil on production performance and fillet lipid and fatty acid composition of juvenile cobia *Rachycentron canadum*. *Aquaculture Nutrition* 17:e437–e447.
- Turchini, G. M., R. M. Gunasekera, and S. S. De Silva. 2003a. Effect of crude oil extracts from trout offal as a replacement for fish oil in the diets of the Australian native fish Murray cod *Maccullochella peelii peelii*. *Aquaculture Research* 34:697–708.
- Turchini, G. M., T. Mentasti, L. Frøyland, E. Orban, F. Caprino, V. M. Moretti, and F. Valfrè. 2003b. Effects of alternative dietary lipid sources on performance, tissue chemical composition, mitochondrial fatty acid oxidation capabilities and sensory characteristics in brown trout (*Salmo trutta* L.). *Aquaculture* 225:251–267.
- Turchini, G. M., T. Mentasti, F. Caprino, S. Panseri, V. M. Moretti, and F. Valfrè. 2004a. Effects of dietary lipid sources on flavour volatile compounds of brown trout (*Salmo trutta* L.) fillet. *Journal of Applied Ichthyology* 20:71–75.
- Turchini, G. M., T. Mentasti, C. Crocco, T. Sala, C. Puzzi, V. M. Moretti, and F. Valfrè. 2004b. Effects of the extensive culture system as finishing production strategy on biometric and chemical parameters in rainbow trout. *Aquaculture Research* 35:378–384.
- Turchini, G. M., D. S. Francis, and S. S. De Silva. 2006. Modification of tissue fatty acid composition in Murray cod (*Maccullochella peelii peelii*, Mitchell) resulting from a shift from vegetable oil diets to a fish oil diet. *Aquaculture Research* 37:570–585.
- Turchini, G. M., D. S. Francis, and S. S. De Silva. 2007. Finishing diets stimulate compensatory growth: results of a study on Murray cod, *Maccullochella peelii peelii*. *Aquaculture Nutrition* 13:351–360.
- Uysal, K., M. Bülbül, M. Dönmez, and A. K. Seçkin. 2008. Changes in some components of the muscle lipids of three freshwater fish species under natural extreme cold and temperate conditions. *Fish Physiology and Biochemistry* 34:455–463.
- Vacha, F., P. Vejsada, J. Huda, and P. Hartvich. 2007. Influence of supplemental cereal feeding on the content and structure of fatty acids during long-lasting storage of common carp (*Cyprinus carpio* L.). *Aquaculture International* 15:321–329.
- Vagner, M., J. H. Robin, J. L. Zambonino Infante, and J. Person-Le Ruyet. 2007a. Combined effects of dietary HUFA level and temperature on sea bass (*Dicentrarchus labrax*) larvae development. *Aquaculture* 266:179–190.
- Vagner, M., J. L. Zambonino Infante, J. H. Robin, and J. Person-Le Ruyet. 2007b. Is it possible to influence European sea bass (*Dicentrarchus labrax*) juvenile metabolism by a nutritional conditioning during larval stage? *Aquaculture* 267:165–174.
- Valente, L. M. P., N. M. Bandarra, A. C. Figueiredo-Silva, A. R. Cordeiro, R. M. Simões, and M. L. Nunes. 2007. Influence of conjugated linoleic acid on growth, lipid composition and hepatic lipogenesis in juvenile European sea bass (*Dicentrarchus labrax*). *Aquaculture* 267:225–235.
- Van Anholt, R. D., W. M. Koven, S. Lutzky, and S. E. Wendelaar Bonga. 2004. Dietary supplementation with arachidonic acid alters the stress response of gilthead seabream (*Sparus aurata*) larvae. *Aquaculture* 238:369–383.
- Van der Meeren, T., A. Mangor-Jensen, and J. Pickova. 2007. The effect of green water and light intensity on survival, growth and lipid composition in Atlantic cod (*Gadus morhua*) during intensive larval rearing. *Aquaculture* 265:206–217.
- Vargas, R. J., S. M. Guimarães de Souza, A. M. Kessler, and S. R. Baggio. 2008. Replacement of fish oil with vegetable oils in diets for jundiá (*Rhamdia quelen* Quoy and Gaimard 1824): effects on performance

- and whole body fatty acid composition. *Aquaculture Research* 39:657–665.
- Vidal, A. T., F. De la Gándara García, A. G. Gómez, and M. J. Cerdá. 2008. Effect of the protein/energy ratio on the growth of Mediterranean yellowtail (*Seriola dumerili*). *Aquaculture Research* 39:1141–1148.
- Villalta, M., A. Estévez, and M. P. Bransden. 2005a. Arachidonic acid enriched live prey induces albinism in Senegal sole (*Solea senegalensis*) larvae. *Aquaculture* 245:193–209.
- Villalta, M., A. Estévez, M. P. Bransden, and J. G. Bell. 2005b. The effect of graded concentrations of dietary DHA on growth, survival and tissue fatty acid profile of Senegal sole (*Solea senegalensis*) larvae during the *Artemia* feeding period. *Aquaculture* 249:353–365.
- Villalta, M., A. Estévez, M. P. Bransden, and J. G. Bell. 2008a. Arachidonic acid, arachidonic/eicosapentaenoic acid ratio, stearidonic acid and eicosanoids are involved in dietary-induced albinism in Senegal sole (*Solea senegalensis*). *Aquaculture Nutrition* 14: 120–128.
- Villalta, M., A. Estévez, M. P. Bransden, and J. G. Bell. 2008b. Effects of dietary eicosapentaenoic acid on growth, survival, pigmentation and fatty acid composition in Senegal sole (*Solea senegalensis*) larvae during the *Artemia* feeding period. *Aquaculture Nutrition* 14:232–241.
- Vizcaino-Ochoa, V., J. P. Lazo, B. Barón-Sevilla, and M. A. Drawbridge. 2010. The effect of dietary docosahexaenoic acid (DHA) on growth, survival and pigmentation of California halibut *Paralichthys californicus* larvae (Ayres, 1810). *Aquaculture* 302:228–234.
- Walker, A. B., H. R. Fournier, C. D. Neefus, G. C. Nardi, and D. L. Berlinsky. 2009. Partial replacement of fish meal with laver *Porphyra* spp. in diets for Atlantic cod. *North American Journal of Aquaculture* 71:39–45.
- Wang, Z., K. Mai, Z. Liufu, H. Ma, W. Xu, Q. Ai, W. Zhang, B. Tan, and X. Wang. 2006. Effect of high dietary intakes of vitamin E and n-3 HUFA on immune responses and resistance to *Edwardsiella tarda* challenge in Japanese flounder (*Paralichthys olivaceus*, Temminck and Schlegel). *Aquaculture Research* 37:681–692.
- Wang, W., L. Hou, X. Zou, F. Yao, B. Yin, and L. Chen. 2007. Effects of n-3 PUFA levels in live foods on albinism, growth, survival, and salinity tolerance of flounder (*Paralichthys olivaceus*) larvae in large-scale artificial rearing. *The Israeli Journal of Aquaculture* 59:137–145.
- Wang, S. Y., K. N. Han, and T. Yoshimatsu. 2008. Comparison between α -LNA and DHA in early developmental stages of *Takifugu obscurus* and *Takifugu rubripes*. *Fisheries Science* 74:853–859.
- Wang, N., S. N. M. Mandiki, E. Henrotte, A.-G. Bouyahia, G. Mairesse, C. Rougeot, C. Melard, and P. Kestemont. 2009. Effect of partial or total replacement of forage fish by a dry diet on the quality of reproduction in pikeperch, *Sander lucioperca*. *Aquaculture Research* 40:376–383.
- Wassef, E. A., N. E. Saleh, and H. A. El-Abd El-Hady. 2009. Vegetable oil blend as alternative lipid resources in diets for gilthead seabream, *Sparus aurata*. *Aquaculture International* 17:421–435.
- Weber, J.-M., G. Brichon, and G. Zwingelstein. 2003. Fatty acid metabolism in rainbow trout (*Oncorhynchus mykiss*) tissues: differential incorporation of palmitate and oleate. *Canadian Journal of Fisheries and Aquatic Sciences* 60:1281–1288.
- Willey, S., D. A. Bengtson, and M. Harel. 2003. Arachidonic acid requirements in larval summer flounder, *Paralichthys dentatus*. *Aquaculture International* 11:131–149.
- Williams, K. C., B. D. Paterson, C. G. Barlow, A. Ford, and R. Roberts. 2003. Potential of meat meal to replace fish meal in extruded dry diets for barramundi, *Lates calcarifer* (Bloch). II. Organoleptic characteristics and fatty acid composition. *Aquaculture Research* 34:33–42.
- Williams, K. C., C. G. Barlow, L. Rodgers, and C. Agcopra. 2006. Dietary composition manipulation to enhance the performance of juvenile barramundi (*Lates calcarifer* Bloch) reared in cool water. *Aquaculture Research* 37:914–927.
- Wilson, C. M., E. N. Friesen, D. A. Higgs, and A. P. Farrell. 2007. The effect of dietary lipid and protein source on the swimming performance, recovery ability and oxygen consumption of Atlantic salmon (*Salmo salar*). *Aquaculture* 273:687–699.
- Wonnacott, E. J., R. L. Lane, and C. C. Kohler. 2004. Influence of dietary replacement of menhaden oil with canola oil on fatty acid composition of sunshine bass. *North American Journal of Aquaculture* 66:243–250.
- Wu, F. C., Y. Y. Ting, and H. Y. Chen. 2003. Dietary docosahexaenoic acid is more optimal than eicosapentaenoic acid affecting the level of cellular defense responses of the juvenile grouper *Epinephelus malabaricus*. *Fish and Shellfish Immunology* 14:223–238.
- Xu, X. L., P. Fontaine, C. Mélard, and P. Kestemont. 2001. Effects of dietary fat levels on growth, feed efficiency and biochemical compositions of Eurasian perch *Perca fluviatilis*. *Aquaculture International* 9:437–449.
- Xu, H., Q. Ai, K. Mai, W. Xu, J. Wang, H. Ma, W. Zhang, X. Wang, and Z. Liufu. 2010. Effects of dietary arachidonic acid on growth performance, survival, immune response and tissue fatty acid composition of juvenile Japanese seabass, *Lateolabrax japonicus*. *Aquaculture* 307:75–82.
- Xu, J.-H., J. Qin, B.-L. Yan, M. Zhu, and G. Luo. 2011. Effects of dietary lipid levels on growth performance, feed utilization and fatty acid composition of juvenile Japanese seabass (*Lateolabrax japonicus*) reared in seawater. *Aquaculture International* 19:79–89.
- Xue, M., L. Luo, X. Wu, Z. Ren, P. Gao, Y. Yu, and G. Pearl. 2006. Effects of six alternative lipid sources on growth and tissue fatty acid composition in

- Japanese sea bass (*Lateolabrax japonicus*). *Aquaculture* 260:206–214.
- Yamamoto, T., K. Teruya, T. Hara, H. Hokazono, H. Hashimoto, N. Suzuki, Y. Iwashita, H. Matsunari, H. Furuita, and K. Mushiake.** 2008. Nutritional evaluation of live food organisms and commercial dry feeds used for seed production of amberjack *Seriola dumerili*. *Fisheries Science* 74: 1096–1108.
- Yamamoto, Y., N. Kabeya, Y. Takeuchi, Alimuddin, Y. Haga, S. Satoh, T. Takeuchi, and G. Yoshizaki.** 2010. Cloning and nutritional regulation of polyunsaturated fatty acid desaturase and elongase of a marine teleost, the nibe croaker *Nibea mitsukurii*. *Fisheries Science* 76:463–472.
- Yasmin, A., T. Takeuchi, M. Hayashi, T. Hirota, W. Ishizuka, and S. Ishida.** 2004a. Effect of conjugated linoleic and docosahexaenoic acids on growth of juvenile tilapia *Oreochromis niloticus*. *Fisheries Science* 70:473–481.
- Yasmin, A., T. Takeuchi, T. Hirota, and S. Ishida.** 2004b. Effect of conjugated linolenic acid (*cis*-9, *trans*-11, *cis*-13–18:3) on growth performance and lipid composition of fingerling rainbow trout *Oncorhynchus mykiss*. *Fisheries Science* 70:1009–1018.
- Yildirim-Aksoy, M., R. Shelby, C. Lim, and P. H. Klesius.** 2007. Growth performance and proximate and fatty acid compositions of channel catfish, *Ictalurus punctatus*, fed for different duration with a commercial diet supplemented with various levels of menhaden fish oil. *Journal of the World Aquaculture Society* 38:461–474.
- Yildirim-Aksoy, M., C. Lim, R. Shelby, and P. H. Klesius.** 2009. Increasing fish oil levels in commercial diets influences hematological and immunological responses of channel catfish, *Ictalurus punctatus*. *Journal of the World Aquaculture Society* 40:76–86.
- Zakęś, Z., A. Kowalska, K. Demska-Zakęś, G. Jeney, and Z. Jeney.** 2008. Effect of two medicinal herbs (*Astragalus radix* and *Lonicera japonica*) on the growth performance and body composition of juvenile pikeperch (*Sander lucioperca* [L.]). *Aquaculture Research* 39:1149–1160.
- Zakęś, Z., B. Jankowska, S. Jarmołowicz, T. Żmijewski, K. Partyka, and K. Demska-Zakęś.** 2010. Effects of different dietary fatty acids profiles on the growth performance and body composition of juvenile tench (*Tinca tinca* [L.]). *Reviews in Fish Biology and Fisheries* 20:389–401.
- Zareh, G., R. Porgholam, A. Shenavar, A. Jafari, and M. Saifzadeh.** 2006. Quality assessment of various meat processing modes for meat from 2-year-old farmed *Huso huso*. *Journal of Applied Ichthyology* 22:422–426.
- Zhao, F., P. Zhuang, L. Zhang, and Z. Shi.** 2010. Biochemical composition of juvenile cultured vs. wild silver pomfret, *Pampus argenteus*: determining the diet for cultured fish. *Fish Physiology and Biochemistry* 36:1105–1111.
- Zhong, Y., S. P. Lall, and F. Shahidi.** 2008. Effects of dietary oxidized oil and vitamin E on the growth, blood parameters and body composition of juvenile Atlantic cod *Gadus morhua* (Linnaeus 1758). *Aquaculture Research* 39:1647–1657.
- Zhu, P., C. C. Parrish, and J. A. Brown.** 2003. Lipid and amino acid metabolism during early development of Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture International* 11:43–52.